# Journal of Integrated

a methodological journal

OMICS

## **Editors-in-Chief**

Carlos Lodeiro-Espiño

Florentino Fdez-Riverola

Jens Coorssen

Jose-Luís Capelo-Martínez

## **JIOMICS**

## Journal of Integrated OMICS

## **Focus and Scope**

Journal of Integrated OMICS, JIOMICS, provides a forum for the publication of original research papers, preliminary communications, technical notes and critical reviews in all branches of pure and applied "-omics", such as genomics, proteomics, lipidomics, metabolomics or metallomics. The manuscripts must address methodological development. Contributions are evaluated based on established guidelines, including the fundamental nature of the study, scientific novelty, and substantial improvement or advantage over existing technology or method. Original research papers on fundamental studies, and novel sensor and instrumentation development, are especially encouraged. It is expected that improvements will also be demonstrated within the context of (or with regard to) a specific biological question; ability to promote the analysis of molecular mechanisms is of particular interest. Novel or improved applications in areas such as clinical, medicinal and biological chemistry, environmental analysis, pharmacology and materials science and engineering are welcome.

## **Editors-in-Chief**

Carlos Lodeiro-Espiño, University NOVA of Lisbon, Portugal Florentino Fdez-Riverola, University of Vigo, Spain Jens R. Coorssen, Brock University, Ontario, Canada Jose-Luís Capelo-Martínez, University NOVA of Lisbon, Portugal

## Regional editors

ASIA

#### Gary Xiao

Director of Functional Genomics and Proteomics Laboratories at Osteoporosis Research Center, Creighton University Omaha, Nebraska, USA

#### Yogeshwer Shukla

Proteomics laboratory at Indian Institute of Toxicology Research (Council of Scientific and Industrial Research), Lucknow, I

## Europe

## Gilberto Igrejas

University of Trás-os-Montes and Alto Douro, Life Sciences and Environmental School, Centre of Genetics and Biotechnology
Department of Genetics and Biotechnology, 5001-801 Vila Real, Portugal

#### Martin von Bergen

UFZ, Helmholtz-Centre for Environmental Research, Department of Proteomics, Permoserstr. 15, 04318 Leipzig, Germany

#### Jan Ottervald

Research and Development | Innovative Medicines Neuroscience, CNSP iMed Science Södertälje, AstraZeneca, Sweden

## North America, Australia and New Zealand

#### Randen Patterson

Center for Computational Proteomics, The Pennsylvania State University, US

#### Yue G

US Environmental Protection Agency, Research Triangle Park, USA

#### Jens R. Coorssen

Brock University, Ontario, Canada

#### South America

#### Eduardo Alves de Almeida

Depto. de Química e Ciências Ambientais, IBILCE - UNESP, Brazil

#### Marco Aurélio Zezzi Arruda

University of Campinas - Unicamp

#### Carlos H. I. Ramos

Chemistry Institute-UNICAMP, Brazil

## **Associated editors**

#### **AFRICA**

## Saffaj Taouqif

Centre Universitaire Régional d'Interface, Université Sidi Mohamed Ben Abdallah, route d'Imouzzar-Fès, Morocco

#### ASIA

#### Abdul Jaleel A

Rajiv Gandhi Centre for Biotechnology, Thycaud PO, Trivandrum, Kerala, India

#### Ali A. Ensafi

Isfahan University of Technology, Iran

#### **Allison Stelling**

Dresden, Germany

#### Amita Pal

Division of Plant Biology, Bose Institute, Kolkata, India

#### Ashish Gupta

Centre of Biomedical Magnetic Resonance, SGPGIMS Campus, Lucknow, India

#### Canhua Huang

The State Key Laboratory of Biotherapy, West China Hospital, Sichuan University, PR China

#### Chaminda Jayampath Seneviratne

Oral Biosciences, Faculty of Dentistry, University of Hong Kong, Hong Kong

#### Cheolju Lee

Korea Institute of Science and Technology, Seoul, Korea

## Chi Chiu Wang

Department of Obstetrics & Gynaecology, Chinese University of Hong Kong, Hong Kong

## Chii-Shiarng Chen

National Museum of Marine Biology and Aquarium, Checheng, Pingtung, Taiwan

#### Ching-Yu Lin

Institute of Environmental Health, College of Public Health, National Taiwan University, Taipei, Taiwan

## Chantragan Srisomsap

Chulabhorn Research Institute, Bangkok, Thailand

### Chen Han-Min

Department of Life Science, Catholic Fu-Jen University, Taipei, Taiwan

#### David Yew

Chinese University of Hong Kong, Shatin, N.T., Hong Kong

#### Debmalya Barh

Institute of Integrative Omics and Applied Biotechnology (IIOAB), India

#### Dwaipayan Bharadwaj

Genomics & Molecular Medicine Unit, Institute of Genomics & Integrative Biology (CSIR), Mall Road, Delhi, India

#### Eiji Kinoshita

Department of Functional Molecular Science, Graduate School of Biomedical Sciences, Hiroshima University, Japan

#### **Eun Joo Song**

Molecular Recognition Research Center, Korea Institute of Science & Technology, Seoul, Korea

## Fan Chen

Institute of Genetics and Developmental Biology, Chinese Academy of Sciences (CAS), China

## Feng Ge

Institute of Hydrobiology, Chinese Academy of Sciences, China

#### Ganesh Chandra Sahoo

BioMedical Informatics Center of Rajendra Memorial Research Institute of Medical Science (RMRIMS), Patna, India

#### Guangchuang Yu

Institute of Life & Health Engineering, Jinan University, Guangzhou, China

## Gufeng Wang

Department of Chemistry, North Carolina State University, Raleigh, USA

#### Hai-Lei Zheng

School of Life Sciences, Xiamen University, China

#### **Heebal Kim**

Department of Food and Animal Biotechnology of the Seoul National University, Korea

#### Hsin-Yi Wu

Institute of Chemistry, Academia Sinica, Taiwan

#### Hitoshi Iwahashi

Health Research Institute, National Institute of Advanced Industrial Science and Technology (AIST), Japan

#### Hong-Lin Chan

National Tsing-Hua University, Taiwan

Hongving Zhong

College of Chemistry, Central China Normal University, Wuhan, P. R. China

## **Huan-Tsung Chang**

Department of Chemistry, National Taiwan University, Taipei, Taiwan

#### HuaXu

Research Resources Center, University of Illinois, Chicago

#### Hui-Fen Wu

 $\label{eq:conditional} Department of Chemistry, National Sun Yat-Sen University, 70, Lien-Hai Road, 80424, Kaohsiung, Taiwan$ 

## **Hye-Sook Kim**

Faculty of Pharmaceutical Sciences, Graduate School of Medicine, Dentistry and Pharmaceutical Sciences, Okayama University, Japan

#### Hyun Joo An

ChungNam National University, Daejeon, Korea (South)

#### Ibrokhim Abdurakhmonov

Institute of Genetics and Plant experimental Biology Academy of Sciences of Uzbekistan, Uzbekistan

#### Isam Khalaila

Biotechnology Engineering Department, Ben-Gurion University, Israel

## Jagannadham Medicharla

Senior Principal Scientist, CSIR-Centre for Cellular and Molecular Biology, Hyderabad, India

#### Jianghao Sun

Food Composition and Method Development Lab, U.S. Dept. of Agriculture, Agricultural Research Services, Beltsville, USA

## Jong Won Yun

Dept. of Biotechnology, Kyungsan, Kyungbuk 712-714, Republic of Korea

## Juan Emilio Palomares-Rius

Forestry and Forest Products Research Institute, Tsukuba, Japan

#### Jung Min Kim

Liver and Immunology Research Center, Daejeon Oriental Hospital of Daejeon University, Republic of Korea

## Kazuaki Kakehi

School of Pharmacy, Kinki University, Kowakae 3-4-1, Higashi-Osaka, 577-8502, Japan

#### Kazuki Sasaki

Department of Molecular Pharmacology, National Cerebral and Cardiovascular Center, Japan

#### Ke Lan

West China School of Pharmacy, Sichuan University, Chengdu, China

#### Kelvin Leung

Department of Chemistry, Hong Kong Baptist University, Hong Kong

#### Kobra Pourabdollah

Razi Chemistry Research Center (RCRC), Shahreza Branch, Islamic Azad University, Shahreza, Iran

#### Kohji Nagano

Chugai Pharmaceutical Co. Ltd., Japan

#### Koji Ueda

Laboratory for Biomarker Development, Center for Genomic Medicine, RIKEN, Tokyo, Japan

#### Krishnakumar Menon

Amrita Center for Nanosciences and Molecular Medicine, Amrita Institute of Medical Sciences, Kochi, Kerala, India

#### Lakshman Samaranavake

Dean, And Chair of Oral Microbiology, University of Hong Kong, Hong Kong

#### Lal Rai

Molecular Biology Section, Centre of Advanced Study in Botany, Banaras Hindu University, Varanasi-221005, India

#### Lei Zhou

Singapore Eye Research Institute, Singapore

#### Li Jianke

Institute of Apicultural Research, Chinese Academy of Agricultural Science, Beijing, China, HKSAR, PR China

#### Ling Zheng

College of Life Sciences, Wuhan University, China

#### Luk John Moonching

National University of Singapore, Singapore

#### Mahdi Ghasemi-Varnamkhasti

Department of Agricultural Machinery Engineering, Faculty of Agriculture, Shahrekord University, Shahrekord, Iran

## Manjunatha Kini

Department of Biological Sciences, National University of Singapore, Singapore

## Masahiro Sugimoto

Graduate School of Medicine and Faculty of Medicine, Kyoto University Medical Innovation Center, Japan

#### Masaya Miyazaki

National Institute of Advanced Industrial Science and Technology, 807-1 Shuku, Tosu, Saga 841-0052, Japan

#### Ming-Fa Hsieh

Department of Biomedical Engineering, Chung Yuan Christian University, Taiwan

#### Mingfeng Yang

Key Laboratory of Urban Agriculture of Ministry of Agriculture P. R. China Beijing University of Agriculture, China

#### Mo Yang

Interdisciplinary Division of Biomedical Engineering, the Hong Kong Polytechnic University, Hong Kong, China

#### **Mohammed Rahman**

Center of Excellence for Advanced Materials Research (CEAMR), King Abdulaziz University, Jeddah, Saudi Arabia

#### Moganty Rajeswari

Department of Biochemistry, All India Institute of Medical Sciences, Ansari Nagar, New Delhi, India

#### Nam Hoon Cho

Dept. of Pathology, Yonsei University College of Medicine, Korea

## Ningwei Zhao

Life Science & Clinical Medicine Dept.; Shimadzu (China) Co., Ltd

## Pei-Yuan Qian

Division of Life Science, Hong Kong University of Science and Technology, China

#### Peng Zhou

Center of Bioinformatics (COBI), Key Laboratory for NeuroInformation of Ministry of Education (KLNME), University of Electronic Science and Technology of China (UESTC)

#### Poh-Kuan CHONG (Shirly)

National University of Singapore, Singapore

#### Qian Shi

Institutes of Biomedical Sciences, Fudan University, Shanghai, China

#### Qionglin Liang

Tsinghua University, Beijing, China

#### Rakesh Mishra

Centre for Cellular and Molecular Biology, Hyderabad, India

#### Roger Beuerman

Singapore Eye Research Institute, Singapore

#### Sameh Magdeldin Mohamed

Niigata prefecture, Nishi-ku, Terao, Niigata, Japan

#### Sanjay Gupta

Advanced Centre for Treatment, Research and Education in Cancer (ACTREC), Tata Memorial Centre, Kharghar, Navi Mumbai, India

#### Sanjeeva Srivastava

Indian Institute of Technology (IIT) Bombay, India

#### Seiichi Uno

Education and Research Center for Marine Resources and Environment, Faculty of Fisheries, Kagoshima University, Japan

#### Sen-Lin Tang

Biodiversity Research Center, Academia Sinica, Taipei, Taiwan

#### Setsuko Komatsu

National Institute of Crop Science, Japan

## Shaojun Dai

Alkali Soil Natural Environmental Science Center, Key Laboratory of Salinealkali Vegetation Ecology Restoration in Oil Field, Ministry of Education, Northeast Forestry University, P.R. China

## Shipin Tian

Institute of Botany, Chinese Academy of Sciences, China

#### **Songping Liang**

Hunan Normal University, Changsha City, China

#### Steven Shaw

Department of Obstetrics and Gynecology, Chang Gung Memorial Hospital, Linkou, Taiwan

## Suresh Kumar

Department of Applied Chemistry, S. V. National Institute of Technology, Gujarat, India

#### Tadashi Kondo

National Cancer Center Research Institute, Japan

#### **Taesung Park**

National Research Laboratory of Bioinformatics and Biostatistics at the Department of Statistics Seoul National University, Korea

## Toshihide Nishimura

Department of Surgery I, Tokyo Medical University, Tokyo, Japan

## Vishvanath Tiwari

 $Department\ of\ Biochemistry,\ Central\ University\ of\ Rajasthan,\ India$ 

#### Wei Wang

School of Medical Sciences, Edith Cowan University, Perth, Australia

#### Weichuan Yu

Department of Electronic and Computer Engineering and Division of Biomedical Engineering, The Hong Kong University of Science and Technology, Clear Water Bay, Kowloon, Hong Kong, China

#### Wei-dong Zhang

Lab of Natural Products, School of Pharmacy, Second Military Medical University, Shangai, China

## Wenxiong Lin

School of Life Sciences, Fujian Agriculture and Forestry University, China

#### William Chen Wei Ning

School of Chemical and Biomolecular Engineering Nanyang Technological University, Singapore

#### Xiao LiWang

Division of Cardiovascular Diseases, Mayo Clinic, Rochester, MN

## Xiao Zhiqiang

Key Laboratory of Cancer Proteomics of Chinese Ministry of Health, Xiangya Hospital, Central South University, 87 Xiangya Road, Changsha, Hunan 410008, P.R. China

#### **Xiaoping Wang**

Key Laboratory of Molecular Biology & Pathology, State Bureau of Chinese Medicine, China

#### **Xuanxian Peng**

School of Life Sciences, Sun Yat-sen University, Guangzhou, China

#### Yang Liu

Department of Chemistry, Tsinghua University, Beijing, China

#### YasminAhmad

Peptide and Proteomics Division Defence Institute of Physiological and Allied Research (DIPAS), DRDO, Ministry of Defence, Timarpur, Delhi-54, India

#### Yin L

Institute of Microbiology, Chinese Academy of Sciences, Beijing, China

#### Yong Song Gho

Department of Life Science, POSTECH, Pohang, Korea

#### Yoon-E Choi

Chonbuk National University, Iksan-si, South Korea

#### Yoon-Pin Lim

Department of Biochemistry, National University of Singapore, Singapore

#### Young-Gvu Ko

College of Life Sciences and Biotechnology, Korea University, Korea

#### Young-Suk Kim

Department of Food Science and Engineering, College of Engineering, Ewha Womans University, Seoul, Korea

#### Youngsoo Kim

Department of Biomedical Sciences, Seoul National University College of Medicine, Seoul, Republic of Korea

#### Youxiong Que

National Research & Development Center for Sugarcane, China Agriculture Research System(CARS), Fujian Agriculture & Forestry University, Republic of China

## Yu-Chang Tyan

Department of Medical Imaging and Radiological Sciences, Kaohsiung Medical University, Kaohsiung, Taiwan

#### Yu Wang

Department of Pharmacology and Pharmacy, the University of Hong Kong, China

#### Yu Xue

Department of Systems Biology, College of Life Science and Technology Huazhong University of Science and Technology, Wuhan, China

#### Yulan Wang

State Key Laboratory of Magnetic Resonance and Atomic and Molecular Physics, Wuhan Centre for Magnetic Resonance, Wuhan Institute of Physics and Mathematics, The Chinese Academy of Sciences, China

#### Zhengwei Yuan

The key laboratory of health ministry for congenital malformation, Shengjing Hospital, China Medical University

## **Zhiqiang Gao**

Department of Chemistry, National University of Singapore

#### AUSTRALIA AND NEW ZEALAND

#### **Bruno Catimel**

Epithelial laboratory, Ludwig Institute for Cancer Research, Post Office Royal Melbourne Hospital, Australia

#### Daniel Cozzolino

Barley Research Laboratory, School of Agriculture, Food and Wine, University of Adelaide, Australia

#### David Reale

CSIRO Land and Water, Highett, Australia

#### **Emad Kiriakous**

Queensland University of Technology (QUT), Brisbane, Australia

#### Joëlle Coumans-Moens

School of Science and Technology, School of Medicine, University of New England, Australia

#### Marc Wilkins

University of New South Wales, Sydney, Australia

#### Maurizio Ronci

Mawson Institute, University of South Australia, Mawson Lakes, Australia

#### Michelle Hill

University of Queensland, Australia

## Michelle Colgrave

CSIRO Livestock Industries, St Lucia, Australia

#### Nicolas Taylor

ARC Centre of Excellence in Plant Energy Biology & Centre for Comparative Analysis of Biomolecular Networks (CABiN), University of Western Australia, Perth. Australia

#### Peter Hoffmann

Institute for Photonics & Advanced Sensing (IPAS), School of Chemistry and Physics, University of Adelaide, Australia

#### **Stefan Clerens**

Protein Quality &Function, AgResearch Ltd Christchurch, New Zealand

### **Peter Solomon**

Research School of Biology College of Medicine, Biology and Environment, Australian National University, Australia

#### Phoebe Chen

Department of Computer Science and Computer Engineering, La Trobe University, Melbourne, Australia

#### Richard Christopherson

School of Molecular Bioscience, University of Sydney, Australia

## Sham Nair

Department of Biological Sciences, Faculty of Science, Macquarie University, NSW, Australia

#### Sylvia Urban

School of Applied Sciences (Discipline of Applied Chemistry), RMIT University, Melbourne, Victoria, Australia

#### Valerie Wasinger

Bioanalytical Mass Spectrometry Facility, Mark Wainwright Analytical Centre, University of NSW, Australia

## Wujun Ma

Centre for Comparative Genomics, Murdoch University, Australia

#### Yin Xiao

Institute of Health and Biomedical Innovation, Queensland University of Technology, Australia

#### AhmetKoc, PhD

Izmir Institute of Technology, Department of Molecular Biology & Genetics, Urla, İzmir, Turkey

#### Alejandro Gella

Department of Basic Sciences, Neuroscience Laboratory, Faculty of Medicine and Health Sciences, Universitat Internacional de Catalunya,

Sant Cugat del Vallès-08195, Barcelona, Spain

#### Alessandro Pessione

Università degli Studi di Torino, Italy

#### **Alexander Scherl**

Proteomics Core Facility, Faculty of Medicine, University of Geneva, Geneva, Switzerland

#### Alfio Ferlito

ENT Clinic, University of Udine, Italy

#### Almudena Fernández Briera

Dpt. Biochemistry Genetics and Immunology, Faculty of Biology –University of Vigo, Spain

## Alfonsina D'Amato

Politecnico di Milano, Department of Chemistry, Materials and Chemical Engineering "GiulioNatta", Italy

#### Alfred Vertegaal

 $Molecular\ Cell\ Biology,\ Leiden\ University\ Medical\ Center,\ The\ Netherlands$ 

#### Ali Mobasheri

School of Veterinary Medicine and Science, Faculty of Medicine and Health Sciences, University of Nottingham, Sutton Bonington Campus, Sutton Bonington, Leicestershire, United Kingdom

#### Andre Almeida

Instituto de Tecnología Química e Biológica, Universidade Nova de Lisboa, Portugal

#### Andrea Matros

Leibniz Institute of Plant Genetics and Crop Plant Research (IPK-Gatersleben), Gatersleben, Germany

#### Andrei Turtoi

University of Liege, Metastasis Research Laboratory, GIGA-Cancer Bât. B23, Belgium

#### Angelo D'Alessandro

Università degli Studi della Tuscia, Department of Ecological and Biological Sciences, Viterbo, Italy

#### Angelo Izzo

Department of Experimental Pharmacology, University of Naples Federico II, Naples, Italy

#### Antonio Gnoni

Department of Medical Basic Sciences, University of Bari "Aldo Moro", Bari, Italy

## Ana Maria Rodríguez-Piñeiro

Institute of Biomedicine, University of Gothenburg, Sweden

## Ana Varela Coelho

Instituto de Tecnologia Química e Biológica (ITQB) Universidade Nova de Lisboa (UNL), Portugal

#### Anna Maria Timperio

Dipartimento Scienze Ambientali Università della Tuscia Viterbo, Italy

#### André Nogueira Da Costa

Molecular Carcinogenesis Group, Section of Mechanisms of Carcinogenesis International Agency for Research on Cancer - World Health Organization (IARC-WHO), Lyon, France

#### **Andreas Boehm**

Steigerfurtweg 8a, D-97084 Würzburg, Germany

## Andrea Scaloni

Proteomics and Mass Spectrometry Laboratory, ISPAAM, National Research Council, via Argine 1085, 80147 Napoli, Italy

#### Andreas Tholey

Division for Systematic Proteome Research, Institute for Experimental Medicine, Christian-Albrechts-University, Germany

## Angel Manteca

Departamento de Biologia Funcional and IUBA, Facultad de Medicina, Universidad de Oviedo, Spain

## Angel P. Diz

Department of Biochemistry, Genetics and Immunology, Faculty of Biology, University of Vigo, Spain

### Angela Bachi

Mass Spectrometry Unit DIBIT, San Raffaele Scientific Institute, Milano, Italy

#### Angela Chambery

Department of Life Science, Second University of Naples, Italy

#### Anna-Irini Koukkou

University of Ioannina, Department of Chemistry, Biochemistry Laboratory, Greece

#### António Sebastião Rodrigues

Departamento de Genética, Faculdade de Ciências Médicas, Universidade Nova de Lisboa, Portugal

#### Arkadiusz Kosmala

Laboratory of Cytogenetics and Molecular Biology, Institute of Plant Genetics, Polish Academy of Sciences, Poland

#### Arzu Umar

Department of Medical Oncology, Laboratory of Breast Cancer Genomics and Proteomics, Erasmus Medical Center Rotterdam Josephine Nefkens Institute, Rotterdam, The Netherlands

#### Baggerman Geert

ProMeta, Interfacultary Center for Proteomics and Metabolomics, Leuven, Belgium

#### Bart De Spiegeleer

Ghent University, Belgium

#### **Bart Devreese**

Laborartory for Protein Biochemistry and Biomolecular Engineering, Department for Biochemistry and Microbiology, Ghent University, Belgium

#### **Bernard Corfe**

Department of Oncology, University of Sheffield, Royal Hallamshire Hospital, United Kingdom

#### Bernd Thiede

Biotechnology Centre of Oslo, University of Oslo, Blindern, Norway

## Björn Meyer

Institut für Instrumentelle Analytik und Bioanalytik Hochschule Mannheim, Germany

#### Bruno Baudin

Biochemistry Laboratory A, Saint-Antoine Hospital, Hôpitaux Universitaires Est Parisien-APHP, Paris, France

#### Bruno Manadas

Center for Neuroscience and Cell Biology, University of Coimbra, Portugal

#### Cândido Pinto Ricardo

Instituto de Tecnologia Química e Biológica, Universidade Nova de Lisboa, Av. da República-EAN, 2780-157 Oeiras, Portugal

## Carla Pinheiro

Plant Sciences Division, Instituto de Tecnologia Química e Biológica (ITQB), Universidade Nova de Lisboa, Portugal

## Claudia Desiderio

Consiglio Nazionale delle Ricerche, Istituto di Chimica del Riconoscimento Molecolare (UOS Roma), Italy

#### Claudio De Pasquale

SAgA Department, University of Palermo, Italy

#### Carlos Gutiérrez Merino

 $\label{thm:continuous} Dept.\ Biochemistry\ and\ Molecular\ Biology\ University\ of\ Extremadura,\ Badajoz,\ Spain$ 

#### Cecilia Calado

Engineering Faculty Catholic University of Portugal, Rio de Mouro, Portugal

#### Celso Reis

Institute of Molecular Pathology and Immunology of the University of Porto, IPATIMUP, Portugal

#### Celso Vladimiro Cunha

Medical Microbiology Department, Institute of Hygiene and Tropical Medicine, New University of Lisbon, Portugal

#### **Charles Steward**

The Wellcome Trust Sanger Institute, Hinxton, United Kingdom

#### **Chris Goldring**

Department of Pharmacology and Therapeutics, MRC Centre for Drug Safety Science, University of Liverpool, United Kingdom

#### Christian Lindermayr

Institute of Biochemical Plant Pathology, Helmholtz Zentrum München, German Research Center for Environmental Health, Neuherberg, Germany

#### Christiane Fæste

Section for Chemistry and Toxicology Norwegian Veterinary Institute, Oslo, Norway

#### **Christer Wingren**

Department of Immunotechnology, Lund University, Lund, Sweden

### Christophe Cordella

UMR1145 INRA, Laboratoire de Chimie Analytique, Paris, France

#### Christophe Masselon

Laboratoire de Biologie a Grande Echelle (iRTSV/BGE), CEA Grenoble, France

## Cosima Damiana Calvano

Universita' degli Studi di Bari, Dipartimento di Chimica, Bari, Italy

#### **David Cairns**

Section of Oncology and Clinical Research, Leeds Institute of Molecular Medicine, Leeds, UK

#### Daniela Cecconi

Dip. diBiotecnologie, LaboratoriodiProteomica e Spettrometriadi Massa, Universitàdi Verona, Verona, Italy

#### **David Honys**

Laboratory of Pollen Biology, Institute of Experimental Botany ASCR, Czech Republic

## David Sheehan

Dept. Biochemistry, University College Cork (UCC), Ireland

#### **Deborah Penque**

Departamento de Genética, Instituto Nacional de Saúde Dr Ricardo Jorge (INSA, I.P.), Lisboa, Portugal

#### Dilek Battal

Mersin University, Faculty of Pharmacy, Department of Toxicology, Turkey

#### Domenico Garozzo

CNR ICTP, Catania, Italy

## **Ed Dudley**

Institute of Mass Spectrometry, College of Medicine Swansea University, Singleton Park, Swansea, Wales, UK

#### Edoardo Saccenti

University of Amsterdam, Netherlands Metabolomics Centre, The Netherlands

#### Elena Gonzalez

Complutense University of Madrid, Dept. Biochemistry and Molecular Biology IV, Veterinary Faculty, Madrid, Spain

#### Elia Ranzato

Dipartimento di Scienze e Innovazione Tecnologica, DiSIT, University of Piemonte Orientale, Alessandria, Italy

## Elisa Bona

Università del Piemonte Oientale, DISIT, Alessandria, Italy

#### Elke Hammer

Interfaculty Institute for Genetics and Functional Genomics, Ernst-Moritz-Arndt Universität, Germany

#### **Enrica Pessione**

University of Torino, Life Sciences and Systems Biology Department, Torino, Italy

#### Eva Rodríguez Suárez

Proteomics Core Facility - CIC bioGUNE, Parque tecnologico de Bizkaia, Spain

#### Federica Pellati

Department of Life Sciences, University of Modena and Reggio Emilia, Italy

#### Ferdinando Cerciello

Laboratory of Molecular Oncology, Clinic of Oncology, University Hospital Zürich, Switzerland

#### Fernando J. Corrales

Division of Hepatology and Gene Therapy, Proteomics Unit, Center for Applied Medical Research (CIMA), Pamplona, Spain

#### Florian Szabados

Dept. of Medical Microbiology, Ruhr-University Bochum, Germany

#### Francesco Saliu

University of Milano Bicocca, Italy

#### Francisco J Blanco

Platform of Proteomics, Proteo-Red-ISCIII INIBIC-Hospital Universitario A Coruña, Spain

#### Francisco Javier Fernández Acero

Laboratory of Microbiology, Marine and Environmental Sciences Faculty, University of Cádiz, Pol. Río San Pedro s/n, Puerto Real, Cádiz, Spain

#### Francisco Torrens

InstitutUniversitari de CiènciaMolecular, Universitat de València, Spain

#### François Fenaille

CEA, IBiTecS, Service de Pharmacologie et DImmunoanalyse (SPI), France

#### Frederic Silvestre

University of Namur, Belgium

#### Fulvio Magni

Department of Health Science, Monza, Italy

#### Georgios Theodoridis

Department of Chemistry, Aristotle University, Greece

## Germain Rousselet

Laboratoire Réparation et Transcription dans les cellules Souches (LRTS), CEA/DSV/IRCM, Fontenay aux Roses, France

#### German Bou

Servicio de Microbiologia-INIBIC, ComplejoHospitalario Universitario La Coruña, Spain

## Gianfranco Mamone

Proteomic and Biomolecular Mass Spectrometry Centre, Institute of Food Science CNR, Italy

#### Gianfranco Romanazzi

Department of Environmental and Crop Sciences, Marche Polytechnic University, Italy

## Gianluigi Mauriello

Department of Food Science, University of Naples Federico II Naples, Italy

## Giorgio Valentini

Università degli Studi di Milano, Dept. of Computer Science, Italy

## Giuseppe Palmisano

Department of Biochemistry and Molecular Biology

University of Southern Denmark, Odense M, Denmark

#### Helen Gika

Chemical Engineering Department, Aristotle University of Thessaloniki, Greece

## Hugo Miguel Baptista Carreira dos Santos

REQUIMTE-FCT Universidade NOVA de Lisboa, Portugal

#### Ignacio Casal

FunctionalProteomicsLaboratory, Centro de Investigaciones Biológicas (CSIC), Madrid, Spain

#### Ignacio Ortea

European Commission, Joint Research Center, Institute for Reference Materials and Measurements, Geel, Belgium

#### Iñaki Álvarez

Institut de Biotecnologia i Biomedicina Vicent Villar Palasí, Universitat Autònoma de Barcelona, Barcelona

#### Isabel Marcelino

Instituto de Tecnología Química e Biológica, Oeiras, Portugal

#### **Isabel Liste**

Area de Biologia Celular y del<br/>Desarrollo, Instituto de Salud Carlos III, Madrid, Spain

#### Isabelle Fournier

University Lille Nord de France, Fundamental & Applied Biological Mass Spectrometry - EA 4550, Villeneuve d'Asco, France

#### Jacek Z. Kubiak

CNRS UMR 6061, University of Rennes 1, Institute of Genetics and Development of Rennes, Rennes, France

#### Jane Thomas-Oates

Centre of Excellence in Mass Spectrometry and Department of Chemistry, University of York, Heslington, UK

#### **Jatin Burniston**

Muscle Physiology and Proteomics Laboratory, Research Institute for Sport and Exercise Sciences, Liverpool John Moores University, Tom Reilly Building, Liverpool, United Kingdom

## Jean-Paul Issartel

INSERM U836, Grenoble Institut des Neurosciences, La Tronche, France

#### Jens Allmer

Molecular Biology and Genetics, Izmir Institute of Technology, Urla, Izmir, Turkey

#### Jerry Thomas

Tecnology Facility, Department of Biology, University of York, UK

#### Jesús Jorrín Novo

Agricultural and Plant Biochemistry, Proteomics Research Group, Department of Biochemistry and Molecular Biology, Córdoba, Spain

#### Jesus Mateos Martín

Osteoarticular and AgingResearch Lab, ProteomicsUnit INIBIC-Complexo Hospitalario Universitario de A Coruña, A Coruña, Spain

#### Joan Cerdà

Laboratory IRTA, Institute of Marine Sciences (CSIC), Passeigmarítim 37-49, 08003 Barcelona, Spain

Joan Claria

Department of Biochemistry and Molecular Genetics, Hospital Clínic of Barcelona, Spain

## João Rodrigues

Instituto de Higiene e Medicina Tropical, Universidade Nova de Lisboa, Portugal

#### Joaquim ROS

 $Dept.\ Ciencies\ Mediques\ Basiques.\ IRB\ Lleida.\ University\ of\ Lleida,\ Spain$ 

#### Joerg Reinders

AG Proteomics, Institute of Functional Genomics, University Regensburg, Germany

## Johan Palmfeldt

Research Unit for Molecular Medicine, Aarhus University Hospital, Skejby, Aarhus, Denmark

#### Jose Andrés Fernández González

Universidad del Pais Vasco, Facultad de Ciencia y Tecnología, Spain

#### Jose Câmara

University of Madeira, Funchal, Portugal

#### Jose Cremata Alvarez

Department of Carbohydrate Chemistry, Center for Genetic Engineering and Biotechnology, Havana, Cuba

#### Jose Luis Martín-Ventura

IIS-FJD-UAM, Madrid, Spain

#### José Manuel Bautista

Departamento de Bioquímica y Biología Molecular IV, Universidad Complutense de Madrid, Spain

#### Jose Manuel Palma

Departamento de Bioquimica, Biologia Celular y Molecular de Plantas

Estacion Experimental del Zaidin, CSIC, Granada, Spain

#### José Moreira

Danish Center for Translational Breast Cancer Research, Denmark

## Juraj Gregan

Max F. Perutz Laboratories, University of Vienna, Austria

#### Karin Stensiö

Department of Photochemistry and Molecular Science, Ångström laboratory, Uppsala University, Sweden

#### Kathleen Marchal

CMPG/Bioinformatics, Dep Microbial and Molecular Systems, Leuven, Germany

#### **Kay Ohlendieck**

Department of Biology, National University of Ireland, Maynooth, Co. Kildare, Ireland

### Keiryn Bennett

CeMM - Center for Molecular Medicine of the Austrian Academy of Sciences Vienna, Austria

#### **Kjell Sergeant**

Centre de Recherche Public-Gabriel Lippmann, Department 'Environment and Agro-biotechnologies' (EVA), Luxembourg

#### **Konstantinos Kouremenos**

Department of Chemistry, Umea University, Sweden

## Lennart Martens

Department of Medical Protein Research, VIB and Department of Biochemistry, Ghent University, Belgium

### Luis P. Fonseca

Instituto Superior Técnico, Centro de Engenharia Biológica e Química, Institute for Biotechnology and Bioengineering, Lisboa, Portugal

#### Luisa Brito

Laboratório de Microbiologia, Instituto Superior de Agronomia, Tapada da Ajuda, Lisbon, Portugal

## Luisa Mannina

CNR, Istituto di Metodologie Chimiche, Rome, Italy

#### Manuel Avilés Sanchez

Department of Cell Biology and Histology, School of Medicine, University of Murcia, Spain

## Mar Vilanova

Misión Biológica de Galicia, Consejo Superior de Inestigaciones Científicas, Pontevedra, Spain

#### Marcello Donini

ENEA -Casaccia Research Center, UTBIORAD-FARM, Biotechnology Laboratory, Italy

## Marco Lemos

GIRM & ESTM - Polytechnic Institute of Leiria, Peniche, Portugal

## Marcus Mau

King's College London, UK

#### María Álava

Departamento de Bioquimica y Biologia Molecular y Celular, Facultad de Ciencias, Universidad de Zaragoza, Spain

Maria De Angelis

Department of Soil, Plant and Food Science, University of Bari Aldo Moro, Italy

#### María de la Fuente

Legume group, Genetic Resources, Mision Biologica de Galicia-CSIC, Pontevedra, Spain

## Maria M. Malagón

Department of Cell Biology, Physiology and Immunology, IMIBIC, Universidad de Córdoba, Spain

#### Maria Gabriela Rivas

REQUIMTE/CQFB, Departamento de Química, Faculdade de Ciências e Tecnologia, Universidade Nova de Lisboa, Portugal

#### María Maván

INIBIC, LaCoruña, Spain

#### María Páez de la Cadena

Department of Biochemistry, Genetics and Immunology, University of Vigo, Spain

#### Marie Arul

Muséum National Histoire Naturelle, Département RDDM, Plateforme de spectrométrie de masse et de protéomique, Paris, France

#### Marie-Pierre Bousquet

Institut de Pharmacologieet de Biologie Structurale, UPS/CNRS, Tolouse, France

#### Mario Diniz

Dept. Química-REQUIMTE, Faculdade de Ciências e Tecnologia, Universidade Nova de Lisboa, Portugal

Mark Davey

Catholic University of Leuven (KU Leuven), Belgium

#### Marko Radulovic

Institute for Oncology and Radiology, Laboratory of Cancer Cell biology, Belgrade, Serbia

#### Martin Hajduch

Department of Reproduction and Developmental Biology, Institute of Plant Genetics and Biotechnology, Slovak Academy of Sciences, Nitra, Slovakia

#### Martin Kussmann

Faculty of Science, Aarhus University, Aarhus, Denmark

#### Martina Marchetti-Deschmann

Institute of Chemical Technologies and Analytics, Vienna University of Technology, Vienna, Austria

#### Maxence Wisztorski

University Lille 1, Laboratoire de Spectrométrie de Masse Biologique, Fondamentale & Appliquée, Villeneuve d'ascq, France

#### Meri Hovsepyan

Institute of Molecular Biology of Armenian National Academy of Sciences Yerevan, Armenia

#### Michalis Nikolaidis

Department of Physical Education and Sports Science at Serres, Aristotle University of Thessaloniki, Greece

#### Michel Jaquinod

Exploring the Dynamics of Proteomes/Laboratoire Biologie à Grande Echelle, Institut de Recherches en Technologies et Sciences pour le Vivant, Grenoble, France

#### Michel Salzet

Laboratoire de Spectrométrie de Masse Biologique Fondamentale et Appliquée, INSERM, Villeneuve d'Ascq, France

## Miguel Reboiro Jato

Escuela Superior de Ingeniería Informática, Ourense, Spain

#### Moncef Mrabet

Laboratory of Legumes (LL), Centre of Biotechnology of Borj-Cédria (CBBC), Hammam-Lif, Tunisia

#### Mónica Botelho

Centre for the study of animal sciences (CECA)/ICETA, Porto, Portugal

#### Monica Carrera

Institute of Molecular Systems Biology, Zurich, Germany

#### Okay Saydam

Molecular Oncology Laboratory, Division of Neuro-Oncology, Department of Pediatrics Medical University of Vienna, Austria

#### Ola Söderberg

Department of Immunology, Genetics and Pathology, Uppsala University, Sweden

#### Paloma Sánchez-Bel

Dpto. Biología del estrés y Patología vegetal, CEBAS-CSIC, Murcia, Spain

## Pantelis Bagos

Department of Computer Science and Biomedical Informatics, University of Central Greece, Greece

#### Paolo Destefanis

Department of Urology, "San Giovanni Battista - Molinette" Hospital, Turin, Italy

## Pasquale Vito

Università del Sannio, Benevento, Italy

#### **Patrice François**

Genomic Research Laboratory, Service of Infectious Diseases, Department of Internal Medicine. Geneva

## Patrícia Alexandra Curado Quintas Dinis Poeta

University of Trás-os-Montes and Alto Douro (UTAD), School of Agrary and Veterinary Sciences, Veterinary, Science Department, Portugal

#### Paul Cutle

F Hoffman La Roche, Basel, Switzerland

#### Paulo Vale

IPMA - Instituto Português do Mar e da Atmosfera, Lisboa, Portugal

#### Pedro Baptista

Centre for Research in Human Molecular Genetics, Department of LifeSciences, Faculdade de Ciências e Tecnologia, Universidade Nova de Lisboa, Caparica, Portugal

## **Pedro Rodrigues**

Centro de Ciências do Mar do Algarve, CCMAR, Faro, Portugal

#### **Pedro Santos**

CBMA-Centre of Molecular and Environmental Biology, Department of Biology, University of Minho, Braga, Portugal

### Pedro S. Lazo

Departamento de Bioquímica y Biología Molecular, Instituto Universitario de OncologíaDel Principado de Asturias (IUOPA), Universidad de Oviedo, Spain

#### Per Bruheim

Department of Biotechnology, Norwegian University of Science and Technology, Trondheim, Norway

#### Phillip Cash

Division of Applied Medicine, University of Aberdeen, Scotland

#### Philipp Hess

Institut Universitaire Mer et Littoral(CNRS - Université de Nantes - Ifremer), Nantes, France

#### Philippe Castagnone-Sereno

Interactions Biotiques et Sante Vegetale, Sophia Antipolis cedex, France

## Pierscionek Barbara

School of Biomedical Sciences, University of Ulster, Cromore Road, Coleraine, BT52 1SA, United Kingdom

#### Pieter de Lange

DipartimentodiScienzedellaVita, SecondaUniversità degli Studi di Napoli, Caserta, Italy

## Qi Zhu

Dept. Electrical Engineering, ESAT/SCD, Katholieke Universiteit Leuven, Heverlee, Belgium

#### Ralph Fingerhut

University Children's Hospital, Swiss Newborn Screening Laboratory, Children's Research Center, Zürich, Switzerland

#### Ralf Hoffmann

Institute of Bioanalytical Chemistry, Center for Biotechnology and Biomedicine, Faculty of Chemistry and Mineralogy, Leipzig University, Germany

#### Rawi Ramautar

Leiden/Amsterdam Center for Drug Research, Leiden University, The Netherlands

#### Ricardo Gutiérrez Gallego

Bioanalysis Group, Neuropsychopharmacology Program IMIM-Hospital del Mar & Department of Experimental and Health Sciences, University Pompeu Fabra, Spain

## Roman Zubarev

Department of Medical Biochemistry and Biophysics, Karolinska Institutet, Stockholm, Sweden

#### Roque Bru Martinez

Plant Proteomics and Functional Genomics Group, Department of Agrochemistry and Biochemistry, Faculty of Sciences, Alicante University, Spain

#### Rubén Armañanzas

Computational Intelligence Group, Departamento de Inteligencia Artificial, Universidad Politécnica de Madrid, Spain

#### **Ruddy Wattiez**

Department of Proteomics and Microbiology, University of Mons (UMONS), Belgium

#### Rune Matthiesen

Institute of Molecular Pathology and Immunology, University of Porto, Portugal

## Ruth Birner-Gruenberger

Medical University Graz, Austria

#### Sabine Luthje

University of Hamburg, Biocenter Klein Flottbek, Hamburg, Germany

#### Sadin Özdemir

Department of Biology, Faculty of Science and Arts, Siirt University, Turkey

#### Salvador Ventura

Institut de Biotecnologia i de Biomedicina, Universitat Autònoma de Barcelona, Spain

## Sandra Kraljevic-Pavelic

University of Rijeka, Department of Biotechnology, Croatia

#### Sebastian Galuska

Institute of Biochemistry, Faculty of Medicine, Justus-Liebig-University of Giessen, Germany

## Serge Cosnier

Department of Molecular Chemistry, Grenoble university/CNRS, Grenoble, France

## Serhat Döker

Cankiri Karatekin University, Chemistry Department, Cankiri, Turkey

#### Shan He

Centre for Systems Biology, School of Biosciences and School of Computer Science, University of Birmingham, England

#### Silvia Mazzuca

Plan Cell Physiology Laboratory, Department of Ecology, University of Calabria, Italy

#### Simona Martinotti

Dipartimento di Scienze e Innovazione Tecnologica, DiSIT, University of Piemonte Orientale, Alessandria, Italy

## Soile Tapio

Helmholtz Zentrum München, German Research Center for Environmental Health, Institute of Radiation Biology, Neuherberg, Germany

#### Sophia Kossida

Biomedical Research Foundation, Academy of Athens, Department of Biotechnology, Athens, Greece

## Spiros D. Garbis

Biomedical Research Foundation of the Academy of Athens, Center for Basic Research - Division of Biotechnology, Greece

#### **Steeve Thany**

Laboratoire Récepteurs et Canaux Ioniques Membranaires, UFR Science, Université d'Angers, France

#### Stefania Orrù

University if Naples Parthenope, Naples, Italy

#### Stefanie Hauck

Research Unit Protein Science, Helmholtz Center Munich, Neuherberg, Germany

#### Stefano Curcio

Department of Engineering Modeling, Laboratory of Transport Phenomena and Biotechnology University of Calabria, Italy

#### Susana Cristóbal

Department of Clinical and Experimental Medicine Faculty of Health Science Linköping University, Sweden

#### Tâmara García Barrera

Departamento de Química y Ciencia de losMateriales, Facultad de Ciencias Experimentales, Universidad de Huelva, Spain

#### Theodore Alexandrov

University of Bremen, Center for Industrial Mathematics, Germany

#### Thole Züchner

Ultrasensitive Protein Detection Unit, Leipzig University, Center for Biotechnology and Biomedicine, Institute of Bioanalytical Chemistry, Germany

#### Tiziana Bonaldi

Department of Experimental Oncology, European Institute of Oncology, Via Adamello 16, 20139 Milan, Italy

#### **Tomris Ozben**

Akdeniz University Medical Faculty Department of Clinical Biochemistry, Antalya, Turkey

#### **Tsangaris George**

Proteomics Research Unit, Center of Basic Research II Foundation of Biomedical Research of the Academy of Athens, Greece

#### ÜnerKolukisaoglu

Center for Plant Molecular Biology, EberhardKarls University Tübingen, Tübingen, Germany

#### Valeria Bertagnolo

Department of Morphology and Embryology University of Ferrara, Italy

#### Vera Muccilli

 $Dipartimento di Scienze Chimiche, Universit\`a di Catania, Catania, Italy$ 

Veronica Mainini

Dept. Health Science, University of Milano-Bicocca, Faculty of Medicine, Monza (MB), Italy

#### Vicenta Martínez-Zorzano

Department of Biochemistry, Genetics and Immunology

University of Vigo, Spain

#### Virginie Brun

French Atomic Energy Commission and French National Institute for Health and Medical Research, France

#### Vittoria Matafora

Biological Mass Spectrometry Unit, San Raffaele Scientific Institute, Milan, Italy

## Vladislav Khrustalev

Department of General Chemistry, Belarussian, State Medical University, Dzerzinskogo, Minsk, Belarus

#### Xiaozhe Zhang

Department of Medicine, University of Frioburg, Switzerland

#### Yuri van der Burgt

Leiden University Medical Center, Department of Parasitology, The Netherlands

#### SOUTH AMERICA

#### Alessandro Farias

Neuroimmunomodulation Group, department of Genetics, Evolution and Bioagents, University of Campinas - SP – Brazil

#### Alexandra Sawaya

Department of Plant Biology, Institute of Biology, UNICAMP, Campinas, São Paulo, Brazil

#### Andréa P.B. Gollucke

Hexalab/Catholic University of Santos, Brazil

#### Arlindo Moura

Department of Animal Science - College of Agricultural Sciences - Federal University of Ceara, Fortaleza, Brasil

#### **Bruno Lomonte**

Instituto Clodomiro Picado, Universidad de Costa Rica

#### Deborah Schechtman

Department of Biochemistry, Chemistry Institute, University of São Paulo, Brazil

#### Edson Guimarães Lo Turco

São Paulo Federal University, Brasil

#### Elisabeth Schwartz

Department of Physiological Sciences, Institute of Biological Sciences, University of Brasilia, Brazil

#### Fabio Ribeiro Cerqueira

Department of Informatics and NuBio (Research Group for Bioinformatics), University of Vicosa, Brazil

#### Fernando Barbosa

Faculty of Pharmaceutical Sciences of Ribeirão Preto University of São Paulo, Brazil

#### Hugo Eduardo Cerecetto

Grupo de Química Medicinal, Facultad de Química, Universidad de la República, Montevideo, Uruguay

### Luis Pacheco

Institute of Health Sciences, Federal University of Bahia, Salvador, Brazil

#### Mário Hiroyuki Hirata

Laboratório de Biologia Molecular Aplicado ao Diagnóstico, Departamento de Análises Clínicas e Toxicológicas, Faculdade de Ciências Farmacêuticas, Universidade de São Paulo, Brazil

## Jan Schripsema

Grupo Metabolômica, Laboratório de Ciências Quimicas, Universidade Estadual do Norte Fluminense, Campos dos Goytacazes, Brazil

#### Jorg Kobarg

Centro Nacional de Pesquisa em Energia e Materiais, Laboratório Nacional de Biociências, Brazil

## Marcelo Bento Soares

Cancer Biology and Epigenomics Program, Children's Memorial Research Center, Professor of Pediatrics, Northwestern University's Feinberg School of Medicine

#### Mario Palma

Center of Study of Social Insects (CEIS)/Dept. Biology, Institute of Biosciences, University of São Paulo State (UNESP), Rio Claro - SP Brazil

#### Rinaldo Wellerson Pereira

Programa de Pós Graduação em Ciências Genômicas e Biotecnologia, Universidade Católica de Brasília, Brazil

#### Roberto Bobadilla

BioSigma S.A., Santiago de Chile, Chile

## Rossana Arroyo

Department of Infectomic and Molecular Biology, Center of Research and Advanced Studies of the National, Polytechnical Institute (CINVESTAV-IPN), Mexico City, Mexico

#### Rubem Menna Barreto

Laboratorio de Biología Celular, Instituto Oswaldo Cruz, Fundação Oswaldo Cruz, Rio de Janeiro, Brazil

#### Vasco Azevedo

BiologicalSciencesInstitute, Federal University of Minas Gerais, Brazil

#### NORTH AMERICA

#### Adam Vigil

University of California, Irvine, USA

#### **Akeel Baig**

Hoffmann-La Roche Limited, Pharma Research Toronto, Toronto, Ontario, Canada

## Alexander Statnikov

Center for Health Informatics and Bioinformatics, New York University School of Medicine, New York

## Amosy M'Koma

Vanderbilt University School of Medicine, Department of General Surgery, Colon and Rectal Surgery, Nashville, USA

#### Amrita Cheema

Georgetown Lombardi Comprehensive Cancer Center, USA

### Anthony Gramolini

 $Department\ of\ Physiology,\ Faculty\ of\ Medicine,\ University\ of\ Toronto,\ Canada$ 

#### Anas Abdel Rahman

Department of Chemistry, Memorial University of Newfoundland and Labrador St. John's, Canada

#### Christina Ferreira

Purdue University - Aston Laboratories of Mass Spectrometry, Hall for Discovery and Learning Research, West Lafayette, US

## **Christoph Borcher**

Biochemistry & Microbiology, University of Victoria, UVic Genome British Columbia Proteomics Centre, Canada

### Dajana Vuckovic

University of Toronto, Donnelly Centre for Cellular + Biomolecular Research, Canada

#### **David Gibson**

University of Colorado Denver, Anschutz Medical Campus, Division of Endocrinology, Metabolism and Diabetes, Aurora, USA

#### Deyu Xie

Department of Plant Biology, Raleigh, USA

#### **Edgar Jaimes**

University of Alabama at Birmingham, USA

## Eric McLamore

University of Florida, Agricultural & Biological Engineering, Gainesville, USA

#### **Eustache Paramithiotis**

Caprion Proteomics Inc., Montreal, Canada

## FangXiang Wu

University of Saskatchewan, Saskatoon, Canada

#### Fouad Daayf

Department of Plant Science, University of Manitoba, Winnipeg, Manitoba, Canada

#### Haitao Lu

Washington University School of Medicine, Saint Louis, USA

#### Hexin Chen

University of South Carolina, Columbia, USA

#### Hsiao-Ching Liu

232D Polk Hall, Department of Animal Science, North Carolina State University Raleigh, USA

## Hui Zhang

Johns Hopkins University, MD, USA

## Ing-Feng Chang

Institute of Plant Biology, National Taiwan University, Taipei, Taiwan

#### Irwin Kurland

Albert Einstein College of Medicine, Associate Professor, Dept of Medicine, USA

#### Jagjit Yadav

Microbial Pathogenesis and Toxicogenomics, Laboratory, Environmental Genetics and Molecular, Toxicology Division, Department of Environmental Health, University of Cincinnati College of Medicine, Ohio, USA

#### Jianbo Yao

Division of Animal and Nutritional Sciences, USA

#### Jiaxu Li

Department of Biochemistry and Molecular Biology, Mississippi State University, USA

#### Jiping Zhu

Exposure and Biomonitoring Division, Health Canada, Ottawa, Canada

#### Jiri Adamec

Department of Biochemistry & Redox Biology Center, University of Nebraska, Lincoln Nebraska, USA

#### Jiye Ai

University of California, Los Angeles

#### John McLean

Department of Chemistry, Vanderbilt University, Nashville, TN, USA

### Joshua Heazlewood

Lawrence Berkeley National Laboratory, Berkeley, CA, USA

#### Kenneth Yu

Memorial Sloan Kettering Cancer Center, New York, USA

#### Laszlo Prokai

Department of Molecular Biology & Immunology, University of North Texas Health Science Center, Fort Worth, USA

#### Lei Li

University of Virginia, USA

#### **Leonard Foster**

Centre for High-throughput Biology, University of British Columbia, Vancouver, BC, Canada

## Madhulika Gupta

Children's Health Research Institute, University of Western Ontario London, ON, Canada

#### Masaru Miyagi

Case Center for Proteomics and Bioinformatics, Case Western Reserve University, Cleveland, USA

## Michael H.A. Roehrl

Department of Pathology and Laboratory Medicine, Boston Medical Center Boston, USA

#### Ming Zhan

National Institute on Aging, Maryland, USA

#### Nicholas Seyfried

Emory University School of Medicine, Atlanta, USA

#### Olgica Trenchevska

Molecular Biomarkers, Biodesign Institute at Arizona State University, USA

#### Peter Nemes

US Food and Drug Administration (FDA), Silver Spring, USA

#### R. John Solaro

University of Illinois College of Medicine, USA

## Rabih Jabbour

Science Application International Corporation, Maryland, USA

#### Ramesh Katam

Plant Biotechnology Lab, Florida A and M University, FL, USA

## Robert L. Hettich

 $Chemical\ Sciences\ Division, Oak\ Ridge\ National\ Laboratory, Oak\ Ridge, USA$ 

#### Robert Powers

University of Nebraska-Lincoln, Department of Chemistry, USA  $\,$ 

#### Shen S. Hu

UCLA School of Dentistry, Dental Research Institute, UCLA Jonsson Comprehensive Cancer Center, Los Angeles CA, USA

#### Shiva M. Singh

University of Western Ontario, Canada

#### Susan Hester

United Stated Environmental Protection Agency, Durnam, USA

#### Terry D. Cyr

Genomics Laboratories, Centre for Vaccine Evaluation, Biologics and Genetic Therapies Directorate, Health Products and Foods Branch, Health Canada, Ontario, Canada

#### **Thibault Mayor**

Department of Biochemistry and Molecular Biology, Centre for High-Throughput Biology (CHiBi), University of British Columbia, Canada

#### Thomas Conrads

USA

#### **Thomas Kislinger**

Department of Medical Biophysics, University of Toronto, Canada

#### Wan Jin Jahng

Department of Biological Sciences, Michigan Technological University, USA

#### Wavne Zhou

Marine Biology Laboratory, Woods Hole, MA, USA

#### Wei Jia

US Environmental Protection Agency, Research Triangle Park, North Carolina, USA

#### Wei-Jun Qian

Pacific Northwest National Laboratory, USA

#### William A LaFramboise

Department of Pathology, University of Pittsburgh School of Medicine Shadyside Hospital, Pittsburgh, USA

## Xiangjia Min

Center for Applied Chemical Biology, Department of Biological Sciences Youngstown State University, USA

#### Xiaoyan Jiang

Senior Scientist, Terry Fox Laboratory, BC Cancer Agency, Vancouver, Canada

#### Xu-Liang Cao

Food Research Division, Bureau of Chemical Safety, Health, Ottawa, Canada

## Xuequn Chen

Department of Molecular & Integrative Physiology, University of Michigan, Ann Arbor, USA

#### Ye Fang

Biochemical Technologies, Science and Technology Division, Corning Incorporated, USA

## Ying Qu

Microdialysis Experts Consultant Service, San Diego, USA

#### Ying Xu

Department of Biochemistry and Molecular Biology, Institute of Bioinformatics, University of Georgia, Life Sciences Building Athens, GA, USA

A methodological Journal

## SPECIAL ISSUE: SELECTED ABSTRACTS OF THE IV INTERNATIONAL CAPARICA CONFERENCE ON URINE OMICS AND TRANSLATIONAL NEPHROLOGY 2019 (URINOMICS 2019)

Urinary exosomal miRNAs from chronic kidney disease (CKD) patients: comparison to exosomes from an RPTEC cell culture model system for CKD	1
Ultrasensitive biomarker assays using plasmonic films and surface enhanced Raman spectroscopy	2
Urinary biomarkers of podocyte dysfunction in patients with chronic glomerulonephritis	3
Non-invasive urinary biomarker candidates of interstitial cystitis	5
A prospective study on prevention of contrast – induced nephropathy in Croatia	6
Hypertension in adult polycystic kidney disease: a narrative review	7
Bioactive lipids in human renal cell lesion and repair: Liquid Chromatography - High Resolution Mass Spectrometry (LC-HRMS) as a tool to investigate new lesion/repair biomarkers	8
Metabolomics study by liquid chromatography - high resolution mass spectrometry (LC-HRMS) to investigate ischemia/ATP depletion injury in Human kidney renal cells: Searching for lesion/repair biomarkers and bioactive lipids	9
Paracetamol sulfonation to investigate sulfotransferase activity in man	10
AKI prediction using NGAL biomarker. A critical point of view on the renal biomarker interpretation	11
Diagnosis of Fluorosis and its Recovery; Fluorosis linked Renal failure	12
First-void urine for detection of cancer biomarkers	14



A METHODOLOGICAL JOURNAL HTTP://www.iiomics.com



SPECIAL ISSUE: SELECTED ABSTRACTS OF THE IV INTERNATIONAL CAPARICA CONFERENCE ON URINE OMICS AND TRANSLATIONAL NEPHROLOGY 2019 (URINOMICS 2019)

# Urinary exosomal miRNAs from chronic kidney disease (CKD) patients: comparison to exosomes from an RPTEC cell culture model system for CKD

Alexander Hüttenhofer<sup>1,3\*</sup>, Glory Ranches<sup>1</sup>, Herbert Schramek<sup>2</sup>, Michael Rudnicki<sup>2</sup>, Gert Mayer<sup>2</sup>

<sup>1</sup> Division of Genomics and RNomics, Biocenter, Medical University Innsbruck, Austria; <sup>2</sup> Department of Internal Medicine IV, Nephrology and Hypertension, Medical University Innsbruck, Innsbruck, Austria; <sup>3</sup> i-med GenomeSeq Core, Innsbruck, Austria

Available Online: 27 December 2019

#### ABSTRACT

Tubular inflammation is a hallmark feature of chronic kidney disease (CKD), an extremely prevalent major public health problem. Understanding the molecular mechanisms in response to tubular inflammation may also pave the way to identify sensitive and efficient diagnostic biomarkers necessary for predicting and treating CKD in the future. In this study, we describe exosomal miRNAs in a cell culture system for CKD as potential regulators and/or biomarkers of CKD. To identify exosomal miRNAs, which are regulated in response to tubular inflammation, we employed several cytokines (IL-1\beta, Oncostatin M and TGF-β1, respectively) to transform renal proximal tubular epithelial cells (RPTECs = CK-) into a wellestablished cell culture model system for CKD (= CK+). The morphology of extracelular vesicles released into the RPTEC culture medium was verified by transmission electron microscopy (TEM) as well as immunoblotting of a tumor susceptibility gene (TSG101), a reported protein marker of exosomes, was performed. By employing a human miRNome PCR panel I, 30 differentially expressed/abundant (p<0.05) miRNAs were identified in exosomes derived from CK+ cells relative to the CK- cells. Among these miRNAs, miR-192-5p, miR- 215-5p, miR-21-5p and miR-23b-5p, respectively, were found to be consistently differentially expressed/abundant (p<0.05) relative to exosomal miRNAs derived from CKcells. Subsequently, these exosomal miRNAs were technically validated and analysed for their release into exosomes. Comparison of the ratio of exosomal miRNAs, relative to the CKD+ or CKD- cells they were derived from, showed an enhanced release (p<0.05) of these exosomal miRNAs for CK+ cells when compared to CK- cells. Urinary exosomes are constitutively released from epithelial cells. In order to assess whether differentially expressed exosomal miRNAs from RPTEC cells resembled exosomal miRNAs released into urine, an identical miRNome PCR panel I analysis was employed from urine samples from healthy controls (n=6) and CKD patients (n=6), respectively. In these analyses, urinary derived exosomal miRNA profiling showed 20 differentially expressed/abundant (p<0.05) miRNAs in CKD patients relative to healthy controls. Interestingly, eight of these urinary exosomal miRNAs overlapped with the differentially expressed RPTEC exosomal miRNAs, suggesting that these miRNAs may be - at least partly - derived from kidney cells and may reflect changes in miRNA abundance in exosomes in response to tubular inflammation. Our study thus provides novel insights into the regulation of potential biomarkers in tubular inflammation in a cell culture system of CKD, and hence these differentially expressed miRNAs may be employed in elucidating the pathophysiological processes in CKD in the future.

Correspondence: Email - alexander.huettenhofer@i-med.ac.at



A METHODOLOGICAL JOURNAL HTTP://www.iiomics.com



SPECIAL ISSUE: SELECTED ABSTRACTS OF THE IV INTERNATIONAL CAPARICA CONFERENCE ON URINE OMICS AND TRANSLATIONAL NEPHROLOGY 2019 (URINOMICS 2019)

# Ultrasensitive biomarker assays using plasmonic films and surface enhanced Raman spectroscopy

Jerry Morrissey

Washington University School of Medicine in St. Louis

Available Online: 27 December 2019

#### ABSTRACT

Fluorescence-based techniques are the cornerstone of modern biomedical optics, with applications ranging from bioimaging at various scales (organelle to organism) to detection and quantification of a wide variety of biological species of interest. However, the weakness of the fluorescence signal remains a persistent challenge in meeting the ever-increasing demand to image, detect, and quantify biological species with low abundance. Here, we report a simple and universal method based on a flexible and conformal elastomeric film with adsorbed plasmonic nanostructures, which we term a "plasmonic patch," that provides large (up to 100-fold) and uniform fluorescence enhancement on a variety of surfaces through simple transfer of the plasmonic patch to the surface. We demonstrate the applications of the plasmonic patch in improving the sensitivity and limit of detection (by more than 100 times) of fluorescence-based immunoassays implemented in microtiter plates and in microarray format. The novel fluorescence enhancement approach presented here represents a disease, biomarker, and application agnostic ubiquitously applicable fundamental and enabling technology to immediately improve the sensitivity of existing analytical methodologies in an easy-to-handle and cost-effective manner, without changing the original procedures of the existing techniques. Using the power of Surface Enhanced Raman Spectroscopy, capture and detection antibodies of a conventional ELISA kit were modified to measure fg/ml amounts of KIM-1 and cytokine IL-6. This sensitivity allows dilution of samples to eliminate the influence of pH and/or solute concentration that would otherwise affect conventional ELISA results. Also, the sensitivity of the assay allows the use of small samples of urine or plasma from infants, small children or from small animals such as mice with experimental remains diseases. It would be possible to detect preclinical changes in KIM-1 or IL-6 prior to disease manifestation. This assay strategy can be adapted to multiplexing to measure two or more analytes simultaneously in one sample by utilizing different Raman reporter molecules on different detection antibodies, an appropriate mix of capture antibodies on the silicon wafer and monitoring nonoverlapping Raman bands specific for each reporter.

Add-on Plasmonic Patch as a Universal Fluorescence Enhancer Light: Sci. Appl.2018, 7, 29. DOI: 10.1038/s41377-018-0027-8

Correspondence: Email - morrisseyjj@wustl.edu



A METHODOLOGICAL JOURNAL HTTP://WWW.JIOMICS.COM



SPECIAL ISSUE: SELECTED ABSTRACTS OF THE IV INTERNATIONAL CAPARICA CONFERENCE ON URINE OMICS AND TRANSLATIONAL NEPHROLOGY 2019 (URINOMICS 2019)

# Urinary biomarkers of podocyte dysfunction in patients with chronic glomerulonephritis

Natalia Chebotareva\*, Irina Bobkova, Vladimir Varshavsky, Olga Li, Lidia Lysenko

Sechenov First Moscow State Medical University (Sechenov University)

Available Online: 27 December 2019

#### ABSTRACT

The role of podocytes in the mechanisms of proteinuria and chronic glomerulonephritis (CGN) progression is the subject of scientific research recently. Biomarkers of podocyte dysfunction can also provide insight into disease progression, prognosis and monitoring of CGN.

Aim: to estimate the podocyte injury markers in patients with chronic glomerulonephritis (CGN).

Methods: 73 CGN pts were studied: 20 - with inactive CGN (I group), 23 active CGN - with proteinuria (PU) > 1 g/d (II group), 30 - with nephrotic syndrome (NS) (III group), including 11 pts with severe NS (PU more than 10 g/d, hypoalbuminemia < 20 g/L)(IIIb), and 19 pts - with moderate NS (IIIa). 8 healthy subjects were studied as control. Podocyturia (PdcU) was estimated by flow cytometry method. The levels of nephrin, heat shock protein-27 (HSP- 27), VEGF, matrix metalloproteinase-2 (MMP-2) and tissue inhibitor of metalloproteinase-2 (TIMP-2) in urine were assessed by ELISA technique. Using immunohistochemistry with anti- WT-1 monoclonal antibody, glomerular podocytes were marked in patients with active glomerulonephritis. The role of urinary markers in predicting response to immunosuppressive therapy was assessed.

The PdcU, NU and HSP-27 urinary levels were higher in active CGN pts than in control group, in group III - significantly higher than in group II (p<0.05) (Tab 1). Podocyturia, NU and urinary HSP-27 level positively correlated with PU (Rs = 0.27, p < 0.05) and negatively with the level of serum albumin (Rs = -0.22, p = 0.07) in pts with active CGN. PdcU and urinary HSP-27 levels were higher in severe nephrotic syndrome compared to moderate NS. Correlation between urinary HSP-27, PdcU and NU levels was found. An imbalance between podocyte markers (PdcU, NU), proinflammatory factors (interleukin-6, MMP-2) and selfdefense factor (VEGF, TIMP-2) was observed in pts with severe NS and renal dysfunction. Urinary VEGF was increased in pts with NS, but reduced in pts with renal dysfunction. The glomerular podocyte number correlated with podocyturia and serum creatinine/GFR. 82% pts with low NU and/or PdcU levels (<17ng/ml and 20/ $\mu$ l respectively) had NS remission within 6 months of active immunosuppressive therapy. On the other hand, 67% pts with high NU and/or PdcU levels (>17ng/ml or 20/ $\mu$ l respectively) showed no significant response to immunosuppressive agents, given from 9 month to 2 years. ROC curve analysis demonstrated the potential role of urine markers as predictors of the response to immunosuppressive therapy in CGN.

Nephrinuria, podocyturia and urinary VEGF excretion may be useful noninvasive tests for assessment of podocyte injury and prognosis of CGN.

## References:

[1] M. Hara, T. Yanagihara, I. Kihara. Clin J Am Soc Nephrol 2 (2007) 231-8.

[2] P. Habara, H. Marecková, Z. Sopková, K. Malícková, D. Zivorová, T. Zima, V. Tesar. Foliav Biol (Praha). 54(5) (2008) 162-7

Correspondence: Email - natasha\_tcheb@mail.ru

Table 1 - Markers of podocyte dysfunction in urine of CGN patients (n=73)

Groups	n	NU, нг/мл	PdcU (pdx+cells/mkl)	VEGF (pg/ml)	HSP-27 ng/ml
Control	8	7.9 [1.7- 9.5]	0[0-0.6]	62.2 [54.85-73.5]	0.73 [00.96]
II	23	9.5 [7.6-13.0] ■	5.45 [2.5 <b>-</b> 8.3] <b>*■</b>	73.5[59.4-90.2]	0.76 [0.68 <b>-</b> 1.14]∎
Illa	19	14.8 [10-26.8] *	8.0 [4.2-15.5]*▲	125.2 [94.6-179.7] *	0.94 [0.71-1.73]
IIIb	11	20.45 [16.3-49.0] *▲	36.5[9.0- 80.3]*▲	54.65 [38.7- 71.5]*▲	1.88 *[0.8-7.2]
I	20	7.25 [5.4-9.8]	5.4[1.0-9.43]*	64.1[53.9-75.6]	0.72 [0.65-0.98]

Median [25th - 75th percentile] p<0,05, \* - vs control, ▲- vs group I, ■- vs group III



A METHODOLOGICAL JOURNAL HTTP://www.jiomics.com



SPECIAL ISSUE: SELECTED ABSTRACTS OF THE IV INTERNATIONAL CAPARICA CONFERENCE ON URINE OMICS AND TRANSLATIONAL NEPHROLOGY 2019 (URINOMICS 2019)

## Non-invasive urinary biomarker candidates of interstitial cystitis

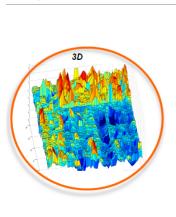
#### Jayoung Kim

Cedars-Sinai Medical Center.

Available Online: 27 December 2019

#### ABSTRACT

Interstitial cystitis/painful bladder syndrome (IC/PBS) is a debilitating condition that presents with a constellation of symptoms including bladder pain, urinary urgency, frequency, nocturia, and small voided volumes in the absence of other identifiable etiologies. A lack of objective diagnostic criteria has affected our ability to adequately treat the disease. The goal of this proposed study is to identify/validate sensitive and non-invasive diagnostic biomarkers using urine specimens that stratify IC/PBS patients from healthy subjects. We performed NMR spectroscopy-based metabolomics analysis to search for soluble metabolites that segregate with the diagnosis of IC/PBS. Annotation of the NMR peaks was performed using MeltDB and MetaboloAnalyst software. We were able to annotate several of the discriminant peaks, including the most significant peak, which was identified as tyramine, a neuro-transmodulator related to pain. These results demonstrate our ability to assay for and provisionally identify discrete urine metabolites that are significantly associated with IC/PBS. We believe this will provide novel insights about the etiology of IC/PBS and identify urine metabolites as biomarkers of IC/PBS that have the potential to be employed clinically.



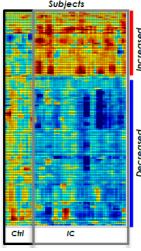


Figure 1 - Identification of NMR Peaks Perturbed in Specimens from IC Patients

#### Acknowledgments:

This work was supported by the National Institutes of Health grants, 1U01DK103260 1R01DK100974, NIH NCATS UCLA CTSI UL1TR000124), Department of Defense grants (W81XWH-15-1-0415), Centers for Disease Controls and Prevention (1U01DP006079), the U.S.-Egypt Science and Technology Joint Fund (to J.K.). The funders had no role in the design, data collection and analysis, decision to publish or preparation of the manuscript. In addition, this article is derived from the Subject Data funded in whole or part by National Academies of Sciences, Engineering and Medicine (NAS) and The United States Agency for International Development (USAID). Any opinions, findings, conclusions, or recommendations expressed in this article are those of the authors alone and do not necessarily reflect the views of USAID or NAS.

Correspondence: Email - jayoung.kim@csmc.edu



A METHODOLOGICAL JOURNAL HTTP://WWW.JIOMICS.COM



SPECIAL ISSUE: SELECTED ABSTRACTS OF THE IV INTERNATIONAL CAPARICA CONFERENCE ON URINE OMICS AND TRANSLATIONAL NEPHROLOGY 2019 (URINOMICS 2019)

# A prospective study on prevention of contrast – induced nephropathy in Croatia

Ana Vujaklija Brajković<sup>1\*</sup>, Marija Križić<sup>2</sup>, Jakša Babel<sup>1</sup>, Mia Rora<sup>1</sup>, Radovan Radonić<sup>1</sup>, Ivan Gornik<sup>3</sup>

1 Department of Intensive Care Medicine, University Hospital Centre Zagreb, Zagreb; 2 Department of Oncology, University Hospital Centre Zagreb, Zagreb; 3 Department of Emergency Medicine, University Hospital Centre Zagreb, Zagreb

Available Online: 27 December 2019

#### ABSTRACT

To explore the protective role of hydration, urine alkalization (Na bicarbonate) and high doses of antioxidant (N-acetylcysteine) in the prevention of CIN. Material and methods: In a prospective, randomized, single-blinded study patients were divided into three groups: 1) peroral hydration, 2) Na bicarbonate infusion and 3) Nacetylcysteine (NAC) plus NaHCO3 infusion. Serum creatinine (SCr), blood urea nitrogen (BUN), and neutrophil gelatinase-associated lipocalin (NGAL) were measured before and 48 hours after the angiography. Mehran score was calculated for each patient. Results: The study included 106 patients. Groups were comparable regarding the baseline characteristics. According to Mehran risk score 70 % of patients had a low risk, 24% medium and 6% high risk score for development of CIN. After the procedure renal function was preserved in all patients regardless of the Mehran risk score (Table 1.). The follow up was completed for 73 patients (68 %). Twenty-two patients (32 %) developed chronic kidney disease. Chronic kidney disease developed in patients with positive history of diabetes and in patients who had higher initial Mehran score. Conclusion: The study showed that patients with preserved renal function are not prone to CIN. Regardless of the protocol used, no case of CIN was observed. Our results indicate that adequate hydration is a key component in maintaining the renal function. Higher Mehran score might be useful in predicting the development of chronic kidney disease.

Table 1 - Laboratory values at the baseline and after the procedure

Variable	Variable All patients Controls (N= 106) (N= 37)			NaHCO3 (N= 40)		NaHCO3+NAC (N= 29)		
Baseline								
Serum creatinine (µmol/mL)	101(88-117.25)	97 (89.5-114.25)	а	102(88.5-117.5)	а	104(86.5-130)	а	
BUN (mmol/L)	6.35(5.1-8.0)	6.6(5.5-7.7)	ab	5.7(4.9-7.3)	a	7.6(6.0-9.7)	b	
Creatinine clearance (ml/min)	71.8(54.9-92.5)	69(55.9-90.1)	а	65.1(52.8-91.1)	a	80.3(68.1-97.6)	а	
NGAL (ng/mL)	8.8 (5.5-16.7)	6.9(4.1-14.0)	а	8.4(4.9-13.5)	a	13.7(8.5-28.4)	b	
Post-procedural								
Serum creatinine (µmol/mL)	103(87.0-121.5)	100(90.0-112.5)	а	106(90.0-123.0)	а	99.0(83.0-127.8)	а	
BUN (mmol/L)	5.8 (4.9-7.6)	6.4(58.8)	а	5.4(4.9-6.5)	b	5.9(4.3-7.6)	ab	
Creatinine clearance (ml/min)	74.7(55.3-97.6)	75.0(60.5-105-2)	а	75.5(51.6-93.7)	a	72.1(58.9-94.5)	а	
NGAL (ng/mL)	11.4(5.4-19.9)	10.6(4.6-19.8)	а	10.8(4.7-16.1)	a	14.0(8.8-33.8)	а	
Difference								
Serum creatinine (µmol/mL)	0.0(-5.0 - 8.0)	1.0(-3.5 - 8.0)	а	1.5(-4.0 - 9.0)	a	-2.5 (-14 – 2.0)	b	
BUN (mmol/L)	0.2(-0.7 - 1.3)	0.0(-0.8 - 0.7)	а	0.1(-0.9 - 0.85)	a	1.9(0.43-2.9)	b	
Creatinine clearance (ml/min)	eatinine clearance (ml/min) -2.0(-19.6–17.3) -7.2(-20.0 –26.4)		а	-0.75(-17.9–15.5)	a	2.1 (-21.0 - 16.3)	а	
NGAL (ng/mL)	0.6(-1.27 - 4.3)	1.4 (-0.2 – 3.6)	а	-0.2 (-1.4 – 3-4)	a	1.3 (2.3 – 6.9)	а	
NGAL (ng/mL) 0.6(-1.27 - 4.3) 1.4 (-0.2 - 3.6) a -0.2 (-1.4 - 3-4) a 1.3 (2.3 - 6.9) a  BUN blood urea nitrogen, NGAL neutrophil gelatinase-associated lipocalin								

Correspondence: Email - avujaklija@gmail.com



A METHODOLOGICAL JOURNAL HTTP://www.jiomics.com



SPECIAL ISSUE: SELECTED ABSTRACTS OF THE IV INTERNATIONAL CAPARICA CONFERENCE ON URINE OMICS AND TRANSLATIONAL NEPHROLOGY 2019 (URINOMICS 2019)

## Hypertension in adult polycystic kidney disease: a narrative review

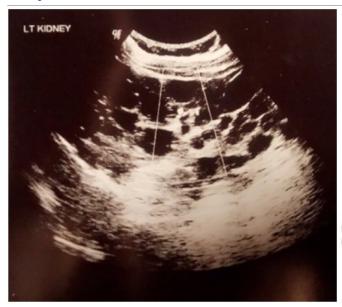
Sarah Mian<sup>1\*</sup>, Yogesh Acharya<sup>2</sup>, Ranjan Dahal<sup>2</sup>

1 Avalon University School of Medicine, Curacao, Netherlands Antilles; 2 Saint Peter's University Hospital, New Jersey, USA.

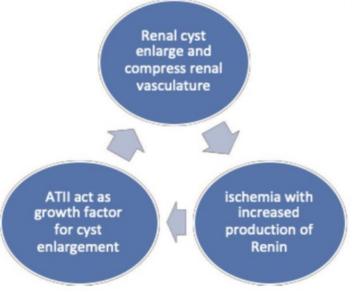
Available Online: 27 December 2019

#### ABSTRACT

Autosomal dominant polycystic kidney disease (ADPKD) is an inherited renal disorder that impacts approximately 12 million worldwide. It is characterized by bilateral kidney enlargement and cystic growth. Hypertension (HTN) is a focal point in the management of ADPKD and is linked to a faster progression to end stage renal disease. Current novel therapies have proven to reduce the progression of renal damage. The ideal goal is to minimize risk through preventative studies and pharmacology to further increase life expectancy and quality. The purpose of this article is to highlight the importance of blood pressure management in ADPKD and review current literature to determine the most effective preventative pharmacotherapy.



**Figure 1** - Ultrasound image showing multiple cysts in kidney.



**Figure 2** - Picture showing vicious cycle ofc yst expansion and subsequent activation of RAAS.

#### References:

- $[1]\ R.\ W.\ Schrier,\ K.\ Z.\ Abebe,\ R.\ D.\ Perrone,\ V.\ E.\ Torres,\ W.\ E.\ Braun,\ T.\ I.\ Steinman,\ et\ al.\ N\ Engl\ J\ Med.\ 371\ (2014)\ 2255-66.$
- [2] F. Rahbari-Oskoui, O. Williams, A. Chapman. Nephrol Dial Transplant. 29 (2014) 2194-201.

Correspondence: Email - sarahamidmian@gmail.com



A METHODOLOGICAL JOURNAL HTTP://WWW.IIOMICS.COM



SPECIAL ISSUE: SELECTED ABSTRACTS OF THE IV INTERNATIONAL CAPARICA CONFERENCE ON URINE OMICS AND TRANSLATIONAL NEPHROLOGY 2019 (URINOMICS 2019)

Bioactive lipids in human renal cell lesion and repair: Liquid Chromatography - High Resolution Mass Spectrometry (LC-HRMS) as a tool to investigate new lesion/repair biomarkers.

Marcelo Einicker-Lamas

Laboratório de Biomembranas, Instituto de Biofísica Carlos Chagas Filho, Universidade Federal do Rio de Janeiro, Brazil.

Available Online: 27 December 2019

#### ABSTRACT

In spite of the great development in the medical procedures and pharmacology, there has been little progress in the treatment of renal failure, which leads to an increasing number of acute and chronic renal patients worldwide. Cell therapy-based protocols as well as nanomedicine initiatives had emerged as potential alternatives. Our group is trying to merge these concepts to achieve an efficient and feasible protocol to minimize renal injury, exploring the versatility of bioactive lipids signaling within he kidney using UHPLC-Q Orbitrap high resolution MS (IC-HRMS), in order to identify early markers for kidney injury as well as renoprotective molecules. Bioactive lipids are an interesting class of mediators that are either important in injury progression or tissue repair. It is well known that some cellular processes are closely regulated by lipid mediators such as diacylglycerol(DAG), phosphatidic acid (PA), lysophosphatidicacid (LPA) and those from the sphingolipid rheostat: ceramide (Cer), sphingosine and sphingosine-1-phosphate(S1P). For example, DAG is associated with phospholipiase C activation during injury/repair; LPA, through LPA1 receptor, is specially associated to the progression of renal fibrosis; while Cer released after activation of sphingomyelinases can be an indicative of tissue damage. In contrast, S1P formation is associated with cell survival and proliferation. Previous results from our group, had demonstrated that the Plasma Membrane Ca\*- ATPase was placed and active in caveolar microdomains [1], being the assembly of these membrane microdomains closely related to the cholesterol amount in the plasma membrane. Therefore, impairment in the cholesterol synthesis would disturb lipid rafts formation and consequently, inhibit the ion transport cited above. These observations led us to postulate that kidney injury would affect he cholesterol content and the bioactive lipids profile, which would disrupt the physiological regulatory network responsible for ions and other solutes homeostasis. Usin

#### **References:**

- [1] G.G. Tortelote, R.H. Valverde, T. Lemos, A. Guilherme, M. Einicker-Lamas, A. Vieyra, FEBS Letters. 576 (2004) 31-35.
- [2] L.S. Sampaio, P.A. da Silva, V.S. Ribeiro, C. Castro-Chaves, L.S. Lara, A. Vieyra, M. Einicker-Lamas, Lipids in Health and Disease 16 (2017) 245-253.
- [3] K.S. Verdoorn, R.S. Lindoso, J. Lowe, L.S. Lara, A. Vieyra, M. Einicker-Lamas, Nephrology Dialysis and Transplantation. 25 (2010) 3867-3874

Correspondence: Email - einicker@biof.ufrj.br



A METHODOLOGICAL JOURNAL HTTP://www.iiomics.com



SPECIAL ISSUE: SELECTED ABSTRACTS OF THE IV INTERNATIONAL CAPARICA CONFERENCE ON URINE OMICS AND TRANSLATIONAL NEPHROLOGY 2019 (URINOMICS 2019)

Metabolomics study by liquid chromatography - high resolution mass spectrometry (LC-HRMS) to investigate ischemia/ATP depletion injury in Human kidney renal cells: Searching for lesion/repair biomarkers and bioactive lipids

Gloria M. R. Soares Grelle<sup>1,2</sup>, Rafael R. H. F. Valverde<sup>1</sup>, Rafael Garrett<sup>2</sup>, Marcelo Einicker-Lamas<sup>1\*</sup>

- <sup>1</sup> Laboratório de Biomembranas, Instituto de Biofísica Carlos Chagas Filho, Universidade Federal do Rio de Janeiro, Brazil;
- <sup>2</sup> Metabolomics Laboratory (LabMeta-LADETEC), Institute of Chemistry, Federal University of Rio de Janeiro

Available Online: 27 December 2019

#### ABSTRACT

Acute renal disease can be triggered by n different causes, such as ischemia. During ischemic injury, the proximal tubule cels are affected, thus impairing kidney function, which led different groups to develop and improve protocols that mimic ischemic injury in vitro to a better understanding of the molecular and cellular events triggered. However, the use of in vitro protocols still raises doubts about the similarity of that lesion provoked in cells with those observed in vivo. Our objective was to analyze and quantify through LC-HRMS, metabolites in the conditioned medium from cultured renal cells either in the control or in the presence of antimycin A, an ATP depletion agent. We aimed to accurately detect different metabolites directed related to ATP depletion injury, such as inosine, xantine and uric acid, as well as, bioactive lipids that would be related either to lesionor repair processes, such as ceramide (Cer), sphingosine-1 phosphate (S1P), and lysophosphatidic acid (LPA). The conditioned medium from human proximal tubule cells (HK-2 strain) cultured both in the control condition and treated with Antimicin A were collected and a metabolomic study using LC-HRMS - UHPLC-Q Orbitrap was performed. Our results clearly confirm the ATP-depletion lesion, as we could detected metabolites that are classically related to the ischemic process, such as: inosine, xanthine, hypoxanthine and uric acid; besides ceramide, a sphingolipid with a direct correlation with apoptosis. S1P and LPA, which are bioactive lipids known to play a role in kidney protection will be also analyzed in the antimycin A-treated cells, as an increase in their content may indicate that these cells are increasing the production of such bioactive lipids in order to face the ATP depleted cells. Among then, we are particularly interested in the endocannabinoids. Further experiments are on the way to quantify the lesion metabolites, as well as a lipidomic analysis either from the conditioned medium or from the human proximal tubule cells in the e

Correspondence: Email - einicker@biof.ufrj.br



A METHODOLOGICAL JOURNAL HTTP://www.jiomics.com



SPECIAL ISSUE: SELECTED ABSTRACTS OF THE IV INTERNATIONAL CAPARICA CONFERENCE ON URINE OMICS AND TRANSLATIONAL NEPHROLOGY 2019 (URINOMICS 2019)

## Paracetamol sulfonation to investigate sulfotransferase activity in man

Natália Marto<sup>1,2\*</sup>, Judit Morello<sup>3</sup>, Alexandra MM Antunes<sup>3</sup>, Emília C Monteiro<sup>1</sup>, Sofia A Pereira<sup>1</sup>

<sup>1</sup> CEDOC - Chronic Diseases Research Centre, NOVA Medical School|Faculdade de Ciências Médicas, Universidade NOVA de Lisboa, Portugal; <sup>2</sup> Department of Internal Medicine, Hospital da Luz, Lisboa, Portugal; <sup>3</sup> Instituto Superior Técnico, Universidade de Lisboa.

Available Online: 27 December 2019

#### ABSTRACT

Cytosolic sulfotransferases (SULT) are important enzymes in Phase II metabolism of several drugs.[1] These enzymes display wide interindividual variability, which is bound to affect efficacy and adverse reactions for drugs metabolized by SULT and underlie drug interactions relevant in clinical practice.[2]

To evaluate intraindividual variability of SULT activity with a phenotyping method that uses paracetamol as probe substrate.

Population and Methods: This study was approved by the Portuguese National Ethics Committee. A population of 36 healthy adults (12 men and 24 women, 12 on oral contraceptives) received 1 g of oral paracetamol on three different occasions. Paracetamol (P) and its metabolites (paracetamol sulfate - PS, paracetamol glucuronate - PG, paracetamol cysteine-S-conjugate - PC and paracetamol mercapturate - PM) were measured in urine using liquid chromatography-high resolution mass spectrometry. SULT activity was measured as the ratio between PS and the sum P+PS+PG (paracetamol sulfonation index – PSI). Differences between groups were tested using ANOVA. Mean PSI was 0.36±0.08. The frequency histogram revealed a normal distribution of PSI in the studied population (Figure 1). Mean intraindividual coefficient of variation of PSI was 15%. There were no significant differences in PSI between men and women, irrespective of oral contraceptive intake.

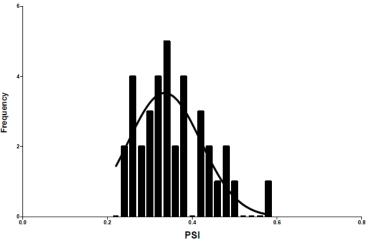


Figure 1 - Frequency histogram plotted as PSI vs. number of individuals.

The use of PSI provides the first evidence of measurements of SULT activity in man. The low intraindividual variability encountered in PSI across genders stimulates further studies envisaging its application in therapeutic drug monitoring and identification of high/poor SULT metabolizers.

#### Acknowledgments:

This study was sponsored by Luz Saude SA.

#### References:

[1] M.W.H. Coughtrie. Chem Biol Interact. 259 (2016) 2-7.

[2] N. Marto, J. Morello, E.C. Monteiro, S.A. Pereira. Drug Metab Rev. 49(3) (2017)357-371.

Correspondence: Email - nfmarto@hospitaldaluz.pt



A METHODOLOGICAL JOURNAL HTTP://www.jiomics.com



SPECIAL ISSUE: SELECTED ABSTRACTS OF THE IV INTERNATIONAL CAPARICA CONFERENCE ON URINE OMICS AND TRANSLATIONAL NEPHROLOGY 2019 (URINOMICS 2019)

# AKI prediction using NGAL biomarker. A critical point of view on the renal biomarker interpretation

Giovanni Introcaso\*, Maria Luisa Biondi

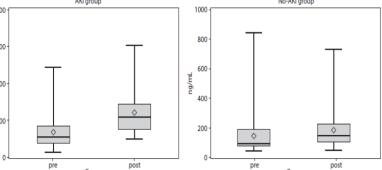
Unit of Laboratory Medicine, Centro Cardiologico Monzino, IRCCS, Milan, Italy

Available Online: 27 December 2019

#### ABSTRACT

In the last years it's well understood that the acute kidney injury (AKI) is increasingly frequent as severe complication after cardiac surgery. An expert opinion suggested to use a biomarker based tubular damage score to identify the cardiac surgery-associated acute kidney injury (CSA-AKI)1. Our work hypothesized that the neutrophil gelatinase-associated lipocalin (NGAL) could be a reliable biomarker to detect early renal impairment. Hence, we conducted a study on 69 patients undergoing cardiac surgery selected by at least two AKI risk factors. Study design provided the NGAL assay by two plasma samples: before surgery (pre NGAL) at general anesthesia induction and within 4 hours from the patient arrival in Intensive Care Unit (ICU) (pos tNGAL), to obtain timely information on acute tubular stress. Serum creatinine (SCrea) was measured every day and after 10·18 hours after surgery to monitor the renal function. According to KDIGO guidelines for AKI definition, a clinical diagnosis was made and patients were divided into AKI group (N= 24) and NO AKI group (N= 45). NGAL interpretation was made considering clinically significant an increase with a second test (post NGAL) 100 ng/mL. Likewise, slight SCrea increases were interpreted in ICU using the reference change value (RCV). Results showed NGAL increases statistically significant only in the AKI group (p< 0,001) (Figure 1).

The better diagnostic outcomes were obtained combining the NGAL increases to the SCrea increases at 10-18 hours post-surgery: sensitivity= 86%, specificity= 70%, NPV= 96%. The combination of NGAL as a tubular damage biomarker to the SCrea as functional marker may be a worthy and efficient strategy to predict AKI in an adult population. In fact, our results may represent a step forward to schedule protocolized biochemical measurements for an early AKI detection then to guide a possible renal replacement therapy.



**Figure 1** - Comparison between AKI and NO AKI groups related to NGAL test pre and postsurgery, continuous variables expressed as mean±SD, evaluated through a t-test for independent samples

#### **Acknowledgments:**

The authors acknowledge the ICU and anesthesiologists staff for the efficient collaboration. A special thank to Dr. Erminio Sisillo as study inspirer and for his fundamental contribution to our work.

## References:

[1] Hilde R.H. de Geus, C. Ronco, M. Haase, L. Jacob, A. Lewington, J.L Vincent. J Thorac Cardiovasc Surg 151 (2016) 1476-81.

Correspondence: Email - giovanni.introcaso@ccfm.it



A METHODOLOGICAL JOURNAL HTTP://WWW.JIOMICS.COM



SPECIAL ISSUE: SELECTED ABSTRACTS OF THE IV INTERNATIONAL CAPARICA CONFERENCE ON URINE OMICS AND TRANSLATIONAL NEPHROLOGY 2019 (URINOMICS 2019)

## Diagnosis of Fluorosis and its Recovery; Fluorosis linked Renal failure

## N. K. Mondal

Fluorosis Foundation of India, B -1, Saransh, 34 – I. P. Extension, New Delhi – 110092, India; Camping and working in Nagaur, Rajasthan, India

Available Online: 27 December 2019

#### EXTENDED ABSTRACT

**Background:** Fluorosis is a public health problem, caused by ingestion of excess fluoride (F) from a variety of sources. Doctors and Clinicians face difficulties in suspecting and diagnosing Fluorosis correctly as the symptoms often overlap with other diseases. The objective of this study is to highlight how to suspect Fluorosis in Out Patient Clinics by retrieving the history and diagnosing the disease through confirmative tests and practice of dietary interventions for recovery. The protocol is well defined and field tested for diagnosis of Fluorosis at an early stage.

Material and Methods: After collecting the patient's history, a battery of tests namely F levels in body fluids (urine & serum) and drinking water are investigated by using Ion Selective Electrode (ISE) potentiometry method besides forearm X-ray radiograph are taken to assess interosseous membrane calcification. Haemoglobin (Hb) is also checked for monitoring purpose. F in all 3 samples tested may be higher compared to normal range with presence of ligamental calcification and this confirmsthe diagnosis of Fluorosis. In patients of Fluorosis, they are advised to practice 2 corrective measures namely diet editing and diet counselling in daily life for recovery. In the former, all sources of F ingestion and use are withdrawn to stop further progression of the disease whereas in the latter, the patient is encouraged to consume a diet rich in essential nutrients, antioxidants and micronutrients through fruits, vegetables and dairy products for repair and maintenance of the damaged body parts. The patients are monitored at intervals to assess F in body fluids and Hb level. Reduction in F levels and rise in Hb have a direct relationship with disappearance of health complaints and subsequent recovery.

Results & discussion: With the history retrieved and the results obtained, a correct diagnosis of Fluorosis is arrived at. For a better understanding of management-cum-monitoring of the patients based on the source of F entry, patients are classified under 3 categories that is summarized in Table 1. Patients were diagnosed Fluorosis with high F in body fluids along with normal or high F level in drinking water besides interosseous membrane calcification in the forearm. The patient was therefore explained the importance and significance of practicing of interventions which focused on elimination F and promotion of nutrients through dietary sources. The main source(s) of F entry identified are (i) drinking of untreated ground water; (ii) consuming F contaminated food / snacks / beverages laced with black rock salt (157 ppm F); (iii) using fluoridated toothpaste The duration of recovery varies from patient to patient depending upon their body physiology and compliance of interventions. It is observed that recovery is faster with nutritive diet than pharmaceutical products. While testing urine and serum fluoride, there is a possibility that results may provide a lead suggestive of renal failure.

**Conclusion:** This communication provides an overview on manifestations of Fluorosis, diagnostic tests, results, differential diagnosis, interventions practiced, monitoring and recovery from the disease. The report highlights that kidney failure may occur due to fluoride toxicity and reveals that it is a Fluorosis linked disorder.

Key words: Fluorosis, Diagnosis, Interventions, Monitoring, Recovery, Case studies, Renal failure.

Correspondence: Email - nisithkm@gmail.com

Table 1 - Comparison Patients classified into 3 categories and interpretation of the test results

	F level in					
	Urine	Serum	Water	X-ray radio- graph showed IMC*	Additional Test	Interpretation
Category 1	<b>↑</b>	<b>↑</b>	<b>↑</b>	$\sqrt{}$	-	Fluorosis confirmed; Source of F entry either contaminated drinking water / food items / other sources rich in F.
Category 2	<b>↑</b>	<b>↑</b>	$\downarrow$	$\checkmark$	-	Fluorosis confirmed; Source of F entry through consumption of food / beverages / other sources rich in F but not drinking water.
Category 3	$\downarrow$	<b>↑</b>	<b>↑</b>	$\sqrt{}$	**KFT	Fluorosis with Renal failure confirmed; Source of F entry either contaminated drinking water/ food items / other sources rich in F.

Normal Range of F: (Urine = 0.1 - 1.0 mg/L); (Serum = 0.02 - 0.05 mg/L); (Drinking Water: Permissible limit for F = Up to 1.0 mg/L, less the better as per BIS, 2012).

## Acknowledgments:

I would like to thank Professor A. K. Susheela, Executive Director, Fluorosis Foundation of India, New Delhi for her constant support and endless guidance during diagnosis of Fluorosis.

## References:

- [1] N. K. Mondal. FLUORIDE 2018; 51(3): 230-242.
- [2] A. K. Susheela, Gupta Rashmi, N. K. Mondal. The National Medical Journal of India 2016; 29 (4); 200-4.
- [3] A. K. Susheela, N. K. Mondal, Tripathi Nalini, Gupta Rashmi. Journal of the Association of Physicians of India 2014; 62:564-71.
- [4] A. K. Susheela, N. K. Mondal, A. Singh. Int J Occup Environ Med 2013; 4:61-72.
- [5] A. K. Susheela, Treatise on Fluorosis. 3rd edition. (ed. Susheela, A.K.), printed by the Fluorosis Research & Rural Development Foundation, New Delhi, 2007.

<sup>\*</sup>IMC - Interosseous Membrane Calcification

<sup>\*\*</sup> KFT – Kidney Function Test



A METHODOLOGICAL JOURNAL HTTP://WWW.JIOMICS.COM



SPECIAL ISSUE: SELECTED ABSTRACTS OF THE IV INTERNATIONAL CAPARICA CONFERENCE ON URINE OMICS AND TRANSLATIONAL NEPHROLOGY 2019 (URINOMICS 2019)

## First-void urine for detection of cancer biomarkers

Quinten Van Avondt\*, Stephanie Jordaens, Arya Mehta, Danielle Pasmans, Koen Beyers, Vanessa Vankerckhoven.

Novosanis NV, Wijnegem, Belgium

Available Online: 27 December 2019

#### EXTENDED ABSTRACT

The predicted global cancer burden is expected to increase significantly – By 2040, 29.5 million new cancer cases are estimated, compared to 18.1 million cases reported worldwide in 2018 [1]. Early detection of cancer can greatly increase chances of survival and improve overall quality of life of a patient. Cancer biomarkers\* can be found in tissues as well as bodily fluids and can be used to detect the disease early as well as monitor disease progression [2]. An ideal biomarker should only be detectable incase of disease and thus have a high sensitivity and specificity. Additionally, biomarker levels can help understand the severity of disease through predictive and prognostic values [3]. A high number of potentially informative cancer biomarkers have been found, based on detection of DNA, RNA, proteins and metabolites. This has been made possible through the ability to sequence the entire human genome as well as advances in key technologies such as high throughput DNA sequencing, microarrays, and mass spectrometry [2].

A tissue biopsy is the traditional approach used to diagnose many cancers. Moreover, biopsies are essential to achieve the objectives of precision oncology and allow for targeted therapies based on the genetic profile of the disease. However, obtaining a tissue sample is not always feasible and the process can be invasive, painful, expensive, time intensive, difficult and requires the intervention of a clinician [4-6]. In addition, due to intratumor heterogeneity, in some instances, the entire tumor landscape may not be reflected by a tissue biopsy. As a result, researchers are continuously exploring alternative methods to detect cancer types. The use of minimally invasive procedures such as liquid biopsies and detection of circulating tumor markers in body fluids is gaining interest [6]. Circulating molecules such as cell-free DNA, circulating tumor cells, circulating RNAs, proteins, peptides and exosomes can provide a global view of primary and metastatic tumors. Circulating molecules can be detected in various biological fluids, including cerebrospinal fluid, plasma, saliva, seminal plasma, serum and urine. Liquid biopsies have several advantages - they allow (repeated) sampling, providing a personalized snapshot of a disease at successive time points. Additionally, they can offer a solution to tumor heterogeneity, and better reflect the genetic profile of all tumor subclones as opposed to tissue biopsies which are obtained from one tumor region. Liquid biopsies are also associated with significantly ess morbidity and can prevent complications associated with traditional biopsies [6].

In general, liquid biopsy is associated with blood, which uses either serum or plasma as a sample type. However, blood as a liquid biopsy has several limitations that have hampered its development as a clinically useful biomarker test. Blood has a relatively high and complex protein repertoire. Furthermore, components of the blood matrix can interfere with biomarker measurements. The invasive nature of blood tests also limits access to repeated measurements and poses a risk of infection for both the patient and caregivers, along with the additional costs of minimizing this risk [7].

Urine has been proposed as an alternative biofluid for detecting and monitoring treatment of urological and systemic cancers. Urine is easily accessible, non-invasive, available in larger quantities and suited for home collection [7]. Moreover, the collection of urine is not limited by the health status of a patient8 and does not entail any risk of transmission of blood-borne pathogens9. In addition, urine testing enables cost-efficient rapid and serial sampling, allowing for patient monitoring as well as for reproducibility assessment of assays [7,8]. In terms of analysis, the isolation of DNA from urine is in theory easier than bbod, due to the low protein content after filtration in the kidney [7,10]. Several studies have shown that the use of urine as a lquid biopsy for cancer detection and monitoring is promising due to the ease of sampling and high acceptability compared to blood

Correspondence: Email - quinten.vanavondt@novosanis.com

and tissue [6,11-13]. Urine cell free tumor DNA has proven to be of value in biomarker studies of bladder, kidney and prostate cancer, but surprisingly also in breast, colon and lung cancer [12, 1416].

However, just as in the case of blood, urine sampling comes with challenges. Collection, transport and storage bring preanalytical variation to the diagnostic process. For example, the amount and proportions of biomarkers in urine might vary according to time of collection and environmental aspects

such as diet. Moreover, urine volume and fraction are preanalytical variables that influence biomarker levels. More specifically, the urine flow breaks up in two distinct fractions. First void urine (FVU) refers to the initial flush of urine, typically the first 10 to 30 mL, in contrast to midstream urine which is considered a sterile sample [17]. FVU contains a higher concentration of biomarkers than other fractions of urine. Many of these preanalytical variables can and should be minimized to improve assay performance. Volumetric and standardized collection is virtually impossible with a urine cup because interrupting urine flow immediately after starting urination is challenging. As a result, sample dilution is likely to occur, and assay sensitivity will be compromised. Additionally, use of a urine cup can be awkward, messy and inconvenient for the user.

To decrease the variation in collected volumes of FVU and to ensure immediate mixing of the sample with a preservative, a sefsampling device has been developed, Colli-Pee®, which allows for standardized and volumetric collection of FVU [11,17]. Colli-Pee is a user-friendly method to capture FVU improving sample collection for downstream analysis, improving diagnostic sensitivity [17,18].

Several diagnostic assays have been developed and/or validated on FVU in the field of oncology. In the case of HPV (Human Papilloma Virus), the primary cause of cervical cancer, testing has traditionally been performed on cervicovaginal samples. As there is a high correlation between urinary and cervical HPV DNA and as it has been shown that FVU contains significantly more human DNA and HPV DNA than the subsequent fractions, the role of urinary detection of HPV in cervical cancer screening is being investigated [11,17]. Leeman et al. showed that FVU samples appear to be suitable for detection of moderate to severe precancerous cervical lesions through HPV testing with high analytical and clinical sensitivity. FVU samples demonstrated high concordance with HPV detected in clinician-taken smears and brush-based self-samples [19]. Moreover, a recent study has investigated the potential of urine-based DNA methylation testing in cervical cancer triage [20]. The drawback of hrHPV (high risk HPV) screening is that it doesn't allow differentiation between a transient productive infection and a persistent transforming infection, which lowers the specificity of the test. Testing of hrHPV-positive women for hypermethylated genes offers an objective triage tool for the detection of CIN3 and cervical cancer [20-22].

With regard to prostate cancer, blood-based PSA testing, characterized by low clinical specificity, is being complemented by novel molecular tests using FVU as a sample. Theodorescu et al. found that FVU testing identified patients with prostate cancer with 91% sensitivity and 69% specificity whereas midstream urine was uninformative [16]. Tests developed by diagnostic companies such as MDxHealth (SelectMDx"), ExosomeDx (IntelliScore®) or Diagnolita measure specific RNA markers for prostate cancer. These tests all enable discrimination between patients with high grade tumors versus patients with low-grade or benign disease at biopsy, reducing over-detection and over treatment of indolent disease.

In conclusion, FVU is a non-invasive sample that is rich in disease biomarkers offering huge potential for infectious disease and cancer biomarker testing. Several diagnostic assays are commercially available or on the verge of launch in the clinical practice. Moreover, research on urine-based liquid biopsy will flourish in the coming years.

#### Acknowledgments:

We would like to thank all participants of the different clinical trials as well as VLAIO for their financial support

## **References:**

- [1] F. Bray, J. Ferlay, I. Soerjomataram, R. L. Siegel, L. A. Torre, A. Jemal. CA [11] J. Pattyn, S. Van Keer, S. Biesmans, et al. J Virol Methods. 264 (2019) 23-Cancer J Clin. 68(6) (2018) 394-424.
- [2] A. K. Gupta, V. P. Reddy, M. Lavania, et al. Indian J Med Res. 132 (2010) [12] Z. Liu, W. Liu. Clin Transl Oncol. 20(8) (2018) 1053-1060. 176-188.
- [3] E. Badila, C. Japie, D. Bartos. Rom J Intern Med. 52(4) (2014) 223-232.
- [4] P. Paudyal, C. Llewellyn, J. Lau, M. Mahmud, H. Smith. PLoS One. 10(4) [15] N. Krishnamurthy, E. Spencer, A. Torkamani, L. Nicholson. J Clin Med. 6 (2015) e0124310.
- [5] S. L. Shih, A. S. Graseck, G. M. Secura, J. F. Peipert. Curr Opin Infect Dis. [16] D. Theodorescu, E. Schiffer, H. W. Bauer, et al. Proteomics Clin Appl. 2 24(1) (2011)78-84.
- [6] A. Di Meo, J. Bartlett, Y. Cheng, M. D. Pasic, G. M. Yousef. Mol Cancer. 16 [17] A. Vorsters, J. Van den Bergh, I. Micalessi, et al. Eur J Clin Microbiol (1) (2017) 80.
- [7] B. M. Nolen, A. E. Lokshin. Int J Biol Markers. 26(3) (2011)141-152.
- [8] A. Franovic, V. M. Raymond, M. G. Erlander, K. L. Reckamp. J Thorac Dis. 9(Suppl 13) (2017) S1323-S1331.
- [9] M. A. Dudeck, J. R. Edwards, K. Allen-Bridson, et al. Am J Infect Control. 43(3) (2015) 206-221.
- [10] Y. H. Su, M. Wang, D. E. Brenner, et al. J Mol Diagn. 6(2) (2004) 101- [22] R. Luttmer, et al. Br. J. Cancer 115 (2016) 579-587.

- [13] D. Xiao, Y. Zeng, S. V. Singh. Mol Carcinog. 48(11) (2009) 1018-1029.
- [14] K Fujita, N. Nonomura. Int J Urol. 25(9) (2018) 770-779.
- (1)(2017).
- (4) (2008) 556-570.
- Infect Dis. 33(11) (2014) 2005-2014.
- [18] D. J. Johnson, A. C. Calderaro, K. A. Roberts. J Forensic Sci. 52(1) (2007) 110 - 113
- [19] A. Leeman, M. Del Pino, A. Molijn, et al. BJOG. 124(9) (2017) 1356-1363.
- [20] B. C. Snoek, A. P. Splunter,, M. C. G. Bleeker, et al. Sci Rep 9 (2019) 3088.
- [21] L. M. A. De Strooper, et al. Cancer Prev. Res. 7 (2014) 1251–1257.